

# Journal Pre-proof

CD34<sup>+</sup> progenitors are predictive of mortality and are associated with physical activity in cardiovascular disease patients

David Muggeridge, Jennifer Dodd, Mark D. Ross



PII: S0021-9150(21)01233-8

DOI: <https://doi.org/10.1016/j.atherosclerosis.2021.07.004>

Reference: ATH 16682

To appear in: *Atherosclerosis*

Received Date: 22 January 2021

Revised Date: 17 June 2021

Accepted Date: 8 July 2021

Please cite this article as: Muggeridge D, Dodd J, Ross MD, CD34<sup>+</sup> progenitors are predictive of mortality and are associated with physical activity in cardiovascular disease patients, *Atherosclerosis* (2021), doi: <https://doi.org/10.1016/j.atherosclerosis.2021.07.004>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

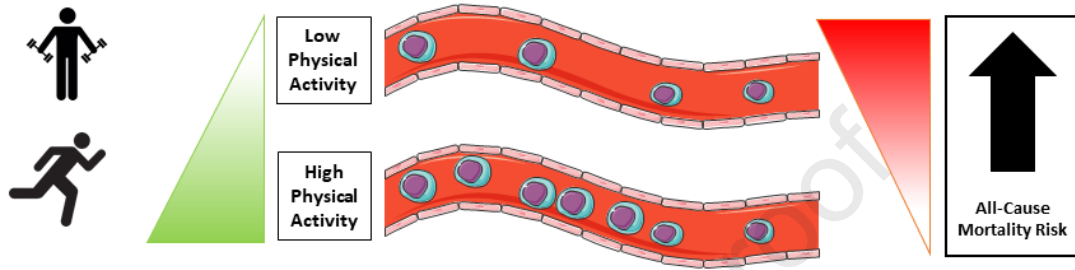
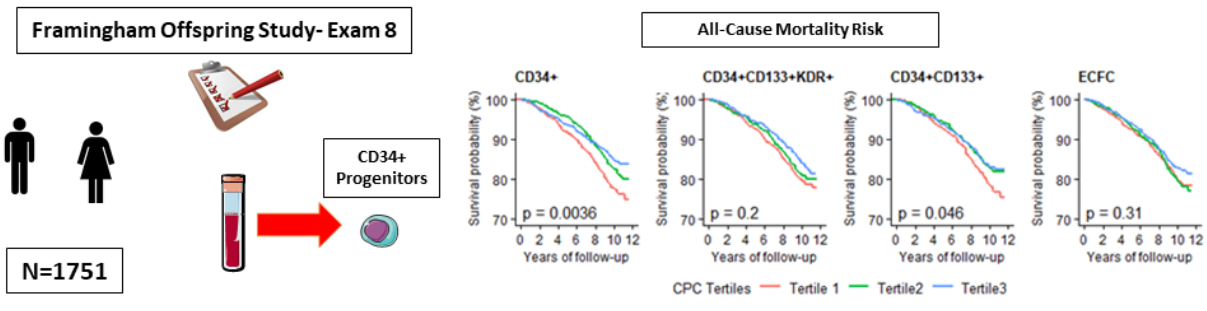
© 2021 Published by Elsevier B.V.

**CD34+ progenitors are predictive of mortality and are associated with physical activity in cardiovascular disease patients**

David Muggeridge<sup>ab</sup> ([D.Muggeridge@napier.ac.uk](mailto:D.Muggeridge@napier.ac.uk)), Jennifer Dodd<sup>a</sup> ([J.Dodd@napier.ac.uk](mailto:J.Dodd@napier.ac.uk)),  
Mark D. Ross<sup>a</sup> ([M.Ross@napier.ac.uk](mailto:M.Ross@napier.ac.uk))

**Credit Author Statement**

M.R conceived and designed the research. D.M, M.R and J.D undertook statistical analysis of the data. M.R and D.M interpreted results of the experiments, prepared figures and drafted the manuscript; all authors edited and revised the manuscript; all authors approved the final version of the manuscript.



Journal Pre-proof

1 **CD34+ progenitors are predictive of mortality and are associated with physical**  
2 **activity in cardiovascular disease patients**

3 David Muggeridge<sup>ab</sup>, Jennifer Dodd<sup>a</sup>, Mark D. Ross<sup>a</sup> ([M.Ross@napier.ac.uk](mailto:M.Ross@napier.ac.uk))

4

5 <sup>a</sup>School of Applied Sciences, Edinburgh Napier University, Edinburgh, United  
6 Kingdom

7 <sup>b</sup>Institute of Health Research & Innovation, Division of Biomedical Science,  
8 University of the Highlands and Islands, Inverness, UK

9

10 **Key Words:** Endothelium, progenitor cells, physical activity, cardiovascular disease,  
11 mortality

12

13 **Corresponding Author:**

14 Mark Ross

15 Edinburgh Napier University

16 School of Applied Sciences

17 Sighthill Campus

18 EH11 4BN

19 [M.Ross@napier.ac.uk](mailto:M.Ross@napier.ac.uk)

20 0131 455 2487

21

22

23

24

25

26 **Abstract**

27

28 *Background and aims:* Circulating progenitor cells (CPCs) play an important role in  
29 vascular repair and can influence cardiovascular (CV) health and longevity. Exercise  
30 is known to modulate these cells via mobilization from the bone marrow. The primary  
31 aims of this study were to evaluate the association of CPCs with mortality and explore  
32 the association between physical activity (PA) and CPCs.

33 *Methods:* 1,751 individuals from the Framingham Offspring cohort ( $66 \pm 9$  years [40-  
34 92 years], 54% female) were included in the study. CPCs ( $CD34^+$ ,  $CD34^+CD133^+$ ,  
35  $CD34^+CD133^+KDR^+$ ) were measured by flow cytometry. Multivariable Cox  
36 regression analyses were performed to investigate relationship of CPCs with future  
37 CV event and mortality. Multivariate regression analyses were performed to  
38 determine the relationship between self-reported PA and CPC counts.

39 *Results:* Following adjustment for standard risk factors, there was an inverse  
40 association between  $CD34^+$  CPCs and all-cause mortality (hazard ratio (HR) per unit  
41 increase in  $CD34^+$ , 0.79; 95% CI 0.64 – 0.98,  $p=0.036$ ).  $CD34^+CD133^+$  CPCs were  
42 inversely associated with CV mortality (HR 0.63, 95% CI 0.44 – 0.91,  $p=0.013$ ).

43 Associations of  $CD34^+$  and  $CD34^+CD133^+$  with mortality were strongest in  
44 participants with pre-existing CVD. PA was associated with  $CD34^+$  CPCs only in  
45 CVD participants (PA Index:  $\beta=0.176$ ,  $p=0.003$ ; moderate-to-vigorous [MVPA]:  
46  $\beta=0.159$ ,  $p=0.007$ ). This relationship was maintained after adjustment for  
47 confounding variables.

48 *Conclusions:* A higher number of  $CD34^+$  and  $CD34^+CD133^+$  CPCs was inversely  
49 associated with all-cause and CV mortality. These associations were strongest in

50 participants with CVD. PA is independently associated with CD34<sup>+</sup> CPCs in  
51 individuals with CVD only, suggestive of greater benefit for this population group.

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

## 75        **1. Introduction**

76

77    Circulating progenitor cells (CPC) are a heterogenous group of cells which have  
78    tissue regenerative potential. A number of studies have shown that CD34<sup>+</sup> CPCs and  
79    several subsets of CD34<sup>+</sup> cells (such as CD34<sup>+</sup>CD133<sup>+</sup>/KDR<sup>+</sup>) can participate in  
80    vascular repair and growth [1–3], and may be associated with vascular endothelial  
81    function [4,5]. Therefore, these cells may reflect vascular integrity and have been  
82    used as biomarkers of vascular repair [6]. CD34<sup>+</sup> CPCs are a diverse group of  
83    progenitors, consisting of both hematopoietic and non-hematopoietic CPCs [7], with  
84    CD133 and KDR often used as more definitive antigen markers for endothelial  
85    progenitor cells (EPC) [8].

86

87    A low number of these CPCs is associated with vascular dysfunction [4,9] and  
88    subsequent greater cardiovascular (CV) risk [10,11]. Observational studies have  
89    shown that individuals with cardiovascular disease (CVD) exhibit lower number and  
90    angiogenic function of these CPCs [12], reflecting reduced vascular repair capacity.  
91    Studies have demonstrated that in individuals hospitalized with heart failure [13], or  
92    with acute coronary syndromes [14], low number of CD34<sup>+</sup> CPCs predicts earlier  
93    mortality in these patients compared to patients with high numbers of CD34<sup>+</sup> CPCs,  
94    which suggests impaired vascular repair capacity in those with higher mortality risk.  
95    Whilst there are no studies that have investigated the role of CD34<sup>+</sup> CPCs and  
96    associated subsets in predicting clinical endpoints in a heterogeneous human  
97    population, there is evidence to suggest that these CPCs are reflective of subclinical  
98    atherosclerotic risk in an apparently healthy population [12].

99

100 Lifestyle behaviors can significantly affect CV health. Smoking [15], physical  
101 inactivity [16] and obesity [17] are associated with perturbed vascular health, leading  
102 to greater risk of mortality. Physical activity, known for its effect on improving  
103 vascular function [18,19] may do so in part via modulating CPC content and/or  
104 function. Studies investigating acute [20–23] and chronic exercise training [24,25]  
105 have demonstrated that progenitor cells can be mobilized into peripheral blood  
106 compartment in humans, where they can exert their vaso-reparative functions.  
107 However, the efficacy of exercise training to promote progenitor cell number has been  
108 argued, with recent evidence demonstrating little or no change in CPC number in  
109 humans after exercise training [6]. As yet, there is no evidence from large cohorts  
110 investigating the association between physical activity and CPCs, with subsequent  
111 patient subgroup (CVD vs CVD-free) analysis to determine if physical activity is  
112 more strongly associated with CPCs in either population.

113

114 The primary aim of this study was to investigate the prognostic potential of CD34<sup>+</sup>  
115 CPCs on all-cause and CV mortality, with the secondary aim to investigate the  
116 relationship between self-reported physical activity on CPCs in a large cohort. It was  
117 hypothesized that circulating CD34<sup>+</sup> CPCs and subpopulations would predict  
118 mortality, and that these cells are associated with self-reported physical activity  
119 levels.

120

121

122

123

124



## 125 2. Materials and methods

126

### 127 2.1 Study sample

128

129 The Framingham Heart Study (FHS) is a longitudinal community-based cohort set up  
130 in 1948 under the direction of the National Heart, Lung, and Blood Institute (NHLBI)  
131 aimed to determine factors that contribute to the onset and progression of  
132 cardiovascular disease (CVD) [26]. Subsequently, an Offspring cohort was included  
133 from 1971 [27]. Participants (n=3,002) in the Framingham Offspring cohort who  
134 attended the 8<sup>th</sup> examination cycle (2004-2008) were eligible for our retrospective  
135 investigation, with n=1,751 included in the study due to availability of key data  
136 (circulating progenitor cells, self-report physical activity levels, follow-up data; see  
137 **Figure 1**). Participant characteristics are shown in **Table 1**.

138

139 This study complies with the Declaration of Helsinki. Ethical approval for all data  
140 collection and research purposes was granted by Boston University Medical Centre,  
141 and written informed consent was obtained for the collection and use of the data  
142 available for secondary investigators. Edinburgh Napier University Research Ethics  
143 and Integrity Committee approved the use of the secondary dataset for the purposes of  
144 the study.

145

### 146 2.2 Clinical assessment

147

148 All participants underwent a clinical and risk factor assessment including assessment  
149 of blood pressure, height and body mass. Fasting blood samples were drawn for

150 quantification of glucose, glycated hemoglobin (HbA1c), total cholesterol, and high  
151 density lipoprotein cholesterol (HDL-C), and triglycerides.

152

### 153 ***2.3 Quantification of circulating progenitor cells***

154

155 Blood samples were collected from participants in the fasted state to quantify CPC  
156 counts. Blood samples were centrifuged and the peripheral blood mononuclear cells  
157 (PBMCs) were isolated for cell phenotyping as previously described [28]. PBMCs  
158 were stained with anti-CD34 FITC, anti-CD133 APC and anti-KDR-PE antibodies  
159 (all BD Biosciences). CD34<sup>+</sup> cells were gated for subsequent expression of CD133  
160 and finally KDR. Total progenitor cells are defined as CD34<sup>+</sup> cells, and EPCs are  
161 defined as CD34<sup>+</sup>CD133<sup>+</sup> and CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup> cells. Analysis of flow cytometry  
162 files were performed using FlowJo analysis software (Treestar, Inc.) and reviewed  
163 by investigators blinded to the identity of the participants.

164

### 165 ***2.4 Endothelial cell colony forming cells (ECFC)***

166

167 In 1653 participants, PBMCs were also used to assess endothelial cell colony forming  
168 cells (ECFC). PBMCs were cultured on fibronectin-coated tissue culture plates (BD  
169 Biosciences) and cultured for 7 days. After 7 days of culture, the number of colonies  
170 in each well was counted by a single blinded individual. ECFC number was reported  
171 as average number of colonies per well up to 12 wells.

172

173

174

## 175 **2.5 Mortality and event incidence**

176

177 Follow up (average:  $9 \pm 2$  years; total: 15,587 person follow-up years) was conducted  
178 for primary end points of all-cause and CV death. Cause of death was determined  
179 through medical history, review of medical records, death certificate, interview of  
180 next of kin, and review of the National Death Index. CV death was defined as death  
181 attributed to ischemic cause (fatal myocardial infarction, stroke). CV event risk was  
182 only assessed in individuals with no pre-existing CVD or CV event occurring before  
183 exam 8 (n = 1467). CV event or incident CVD was assessed using the standard  
184 Framingham Heart Study criteria and included the following: new-onset angina, fatal  
185 and non-fatal MI or stroke, heart failure or intermittent claudication.

186

## 187 **2.6 Self-reported physical activity levels**

188

189 Self-reported sleep, sitting time, light, moderate and heavy activity were determined  
190 using a physical activity questionnaire employed by the Framingham Heart Study.  
191 The number of hours of certain activity per week was collected. A composite score  
192 was calculated (physical activity index; PAI), for each participant by weighting a 24 h  
193 activity recall. Participants were asked to report the number of hours in a typical day  
194 spent sleeping (weighting factor [WF] = 1) and in sedentary (WF = 1.1), slight  
195 (WF = 1.5), moderate (WF = 2.4), and heavy activities (WF = 5) [29]. PAI was  
196 subsequently calculated by adding the products of the hours spent at each activity  
197 domain and their weighting factor based on the oxygen requirements for said activity  
198 [30].

199

## 200 2.7 Statistical analysis

201

202 Continuous variables were assessed for normality by assessing histograms and Q-Q  
203 plots. Data for CD34<sup>+</sup>, CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup>, CD34<sup>+</sup>CD133<sup>+</sup> and PAI were natural  
204 log transformed and EFCFs were square root transformed. Appropriate data  
205 transformations were applied when relevant prior to further statistical analysis.

206 Participants with missing data were excluded and thus complete-case analyses were  
207 performed. All participants were categorized into tertiles for each CPC measure for  
208 event and mortality risk analyses using Kaplan-Meier curve and log-rank analyses.

209 Subsequent Cox proportional hazards regression analyses were performed, utilizing  
210 transformed continuous data for CPC. Cox proportional hazards regressions were  
211 performed unadjusted and adjusted for age, sex, BMI, PAI, CVD and diabetes status,  
212 smoking status. To investigate the effects of CVD status, the data set was split and  
213 analyses repeated for those free of CVD at exam 8 (n = 1467) and those with a CVD  
214 diagnosis prior to exam 8 (n = 284). Proportional hazards assumptions for each of the  
215 Cox models were evaluated by plots of Schoenfeld residuals.

216

217 To assess the influence of physical activity on CPC counts, linear regression analyses  
218 were performed to assess the relationship between CPC counts and PAI. A subset of  
219 physical activity, moderate + heavy activity time, was also investigated. Unadjusted  
220 and adjusted analyses are displayed. Data analyses were carried out using RStudio  
221 Team (2019, RStudio: Integrated Development for R. RStudio, Inc, Boston, MA:  
222 <http://www.rstudio.com/>). *p*-values of <0.05 were considered statistically significant.

223

224

## 225 3. Results

226

### 227 3.1 Relationship between CPC Counts and adverse events

228

#### 229 3.1.1 All-cause mortality

230 Kaplan Meier curves based on tertiles of CPC counts and all-cause mortality are  
231 shown in **Figure 2 A-D**. In unadjusted Cox proportional hazard models, increases in  
232 CD34<sup>+</sup> and CD34<sup>+</sup>CD133<sup>+</sup> CPCs were significantly associated with a decreased risk  
233 of death ( $p < 0.001$ ,  $p = 0.001$ ; **Table 2**). Following adjustment, increases in CD34<sup>+</sup>  
234 remained significantly associated with a decreased risk of death ( $p = 0.036$ ). Whilst  
235 there was a trend for CD34<sup>+</sup>CD133<sup>+</sup> on all-cause mortality, this did not reach  
236 statistical significance ( $p = 0.07$ ). No significant associations were observed for all-  
237 cause mortality for CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup> EPCs or ECFC (all  $p > 0.05$ ; **Table 2**).

238

#### 239 3.1.2 Cardiovascular mortality

240 Kaplan Meier curves based on tertiles of CPC counts and CV mortality are shown in  
241 **Figure 2 E-H**. In unadjusted Cox proportional hazard models, increases in CD34<sup>+</sup>  
242 and CD34<sup>+</sup>CD133<sup>+</sup> CPCs were significantly associated with a decreased risk of CV  
243 death ( $p = 0.008$ ,  $p = 0.006$ ; **Table 2**). Following adjustment, CD34<sup>+</sup>CD133<sup>+</sup> CPCs  
244 were significantly associated with a decreased risk of CV death ( $p = 0.013$ ). Whilst  
245 there was a trend for CD34<sup>+</sup> on CVD mortality, this did not reach statistical  
246 significance ( $p = 0.055$ ). No other significant associations were observed for CVD  
247 mortality (all  $p > 0.05$ ; **Table 2**).

248

249

250

251 **3.2 Relationship between CPC counts and adverse events- influence of CVD status**

252

253 **3.2.1 All-cause mortality**

254 Kaplan Meier curves based on tertiles of CPC counts and all-cause mortality for those

255 free of CVD and those with CVD at exam 8 are shown in **Supplementary Figure 1**.

256 Unadjusted and adjusted Cox proportional hazard models for CPC counts are

257 displayed in **Table 3**. Following adjustment, increases in CD34<sup>+</sup> and CD34<sup>+</sup>CD133<sup>+</sup>

258 CPCs were significantly associated with a decreased risk of death in those with CVD

259 at exam 8 ( $p=0.032$ ,  $p =0.003$ ). No other significant associations were observed for260 all-cause mortality (all  $p>0.05$ ; **Table 3**).

261

262 **3.2.2 Cardiovascular mortality**

263 Kaplan Meier curves based on tertiles of CPC counts and CV mortality for those free

264 of CVD and those with CVD at exam 8 are shown in **Supplementary Figure 1**.

265 Unadjusted and adjusted Cox proportional hazard models for CPC counts and CV

266 mortality are displayed in **Table 3**. In unadjusted and adjusted Cox proportional267 hazard models, increases in CD34<sup>+</sup> and CD34<sup>+</sup>CD133<sup>+</sup> CPCs were significantly

268 associated with a decreased risk of CV mortality in the CVD present at exam 8 group

269 (all  $p<0.05$ , **Table 3**). No other significant associations were observed for CV270 mortality in either of the sub-groups (all  $p>0.05$ ).

271

272 **3.2.3 Cardiovascular events**

273 Cox proportional hazard analysis was performed in the population free of CVD for

274 incidence of future CV events. ECFCs were significantly associated with a decreased

275 risk of future CV events ( $p=0.046$ , **Supplementary Table 1**). There was no  
276 association between CPC counts and CV event risk for all other measures (all  
277  $p>0.05$ ).

278

### 279 *3.3 Association of physical activity with CPC counts*

280

281 To assess the association between physical activity and CPC counts, both unadjusted  
282 and adjusted linear regressions were performed. In unadjusted and adjusted analyses,  
283 PAI and moderate + heavy activity hours were not associated with any CPC subset or  
284 with ECFC units. However, in the CVD group, after adjusting for confounders, both  
285 PAI and moderate + heavy activity time were positively associated with CD34<sup>+</sup> CPCs  
286 and were the only significant predictors of the number of these cells (**Table 4** and  
287 **Supplementary Table 2**). Physical activity was not associated with CD34<sup>+</sup>CD133<sup>+</sup>,  
288 CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup> CPCs or ECFC counts, both in univariate and multivariate  
289 analyses. Light activity time was not significantly associated with CD34<sup>+</sup>,  
290 CD34<sup>+</sup>CD133<sup>+</sup>, CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup> or ECFC counts (all  $p>0.05$ ).

291

## 292 **4. Discussion**

293

294 Our main findings were that CD34<sup>+</sup> and CD34<sup>+</sup>CD133<sup>+</sup> CPCs were significant  
295 predictors of all-cause and CV mortality in the Framingham Offspring cohort, driven  
296 primarily by the strength of this association in individuals with CVD. Additionally,  
297 increase in self-reported physical activity is positively associated with higher CD34<sup>+</sup>  
298 CPCs in our CVD cohort after adjustment for confounders, a relationship not evident  
299 in our CVD-free cohort. Together, these findings suggest that the observed protection

300 of increased CD34<sup>+</sup> CPCs on mortality in a diseased population is partly driven by the  
301 physical activity levels of individuals.

302

303 Several small studies have investigated the prognostic potential of CPCs as  
304 biomarkers of vascular repair for predicting incident risk of all-cause and/or CV  
305 death. These studies have demonstrated that these cells can predict mortality or  
306 clinical end-points in several disease populations, for example patients with coronary  
307 artery disease [31], acute coronary syndromes [14], heart failure [13], or type 2  
308 diabetes [32]. Our data support these observations, with CD34<sup>+</sup> and CD34<sup>+</sup>CD133<sup>+</sup>  
309 CPCs predictive of all-cause and CV mortality. Interestingly, this association was  
310 absent in the CVD-free population and driven mainly by a strong association with  
311 mortality in individuals with pre-diagnosed CVD, suggestive that the prognostic  
312 potential of these cells is much stronger in disease populations, and offers little  
313 predictive potential, if any, in apparently healthy populations. Interestingly,  
314 CD34<sup>+</sup>CD133<sup>+</sup>KDR putative EPCs and ECFCs showed no predictive ability for all-  
315 cause or CV death in our study.

316

317 In the largest study investigating the role of CPCs on incident risk prediction in a  
318 CVD cohort, Patel and colleagues [31] observed that, like our study, only CD34<sup>+</sup> and  
319 CD34<sup>+</sup>CD133<sup>+</sup> CPCs were predictive of mortality. In 2 cohorts, each over 400  
320 patients (n=905 pooled), Patel et al. [31] showed that increases in both these  
321 progenitor subsets showed a significant inverse association with all-cause and CV-  
322 mortality, and that CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup> cells, like our data, showed no association  
323 with mortality, with Werner et al. [33] also demonstrating little prognostic potential  
324 for KDR<sup>+</sup> EPCs on all-cause death, MI and stroke, however, they did show a



325 significant association with CV mortality, which was defined as death from acute MI,  
326 CAD, or congestive heart failure. It is likely that differences in definition of CV death  
327 between these studies may explain the different findings. Our data in >1700  
328 individuals, however, specifically shows that the associations of CD34<sup>+</sup> and  
329 CD34<sup>+</sup>CD133<sup>+</sup> with all-cause and CV mortality are driven by their prognostic  
330 strength in individuals with CVD, and not those who are CVD-free. It is likely that  
331 these cells play a more important role in CVD when the vascular system is in a state  
332 of constant damage, and that a lower number of these cells in these patients reflects  
333 exhaustion of the progenitor cell pool. Interestingly, our data indicated that ECFC  
334 numbers were predictive of future CV event incidence in CVD-free participants,  
335 potentially emphasizing the possibly more sensitive cell culture measures of vascular  
336 repair as opposed to flow cytometric measures. However, ECFCs showed no other  
337 association with all-cause or CV mortality in either population group.

338

339 Both CD34<sup>+</sup> and CD133<sup>+</sup> progenitor cells have vascular regenerative capabilities  
340 [2,34–36]. These cells, reported initially to have pro-angiogenic capabilities due to the  
341 potential to differentiate into endothelial cells [3], most probably work in a paracrine  
342 manner, through secretion of vasoactive and proangiogenic factors, such as VEGF  
343 and other pro-angiogenic cytokines [36]. Due to their potential vasculo-reparative  
344 capacities, clinical studies have been undertaken to assess their efficacy as cellular  
345 therapies to promote recovery of blood flow in myocardial infarction and stroke  
346 studies. Clinical studies showing implantation or injection of these cell types show  
347 promise in repair of damaged myocardium in animal models [1] and in some human  
348 studies [37,38], however, due to the expense and research and development required

349 to optimize this cellular therapy, other non-pharmaceutical interventions may be more  
350 effective in promoting endogenous vascular repair for clinical benefit.

351

352 In addition, given the reduced number [39,40] of CPCs in individuals with CVD, and  
353 the predictive association with mortality [31], it is pertinent to find therapies to  
354 augment production, mobilization and function of these progenitor cells. Exercise and  
355 physical activity have the potential to mobilize CD34<sup>+</sup> cells into the circulation as  
356 evident from acute exercise studies showing transient increases in CPCs in both  
357 healthy [20,21,41,42] and diseased populations [43], although the response to acute  
358 exercise is somewhat diminished in CVD patients [44]. Long-term physical exercise  
359 and physical activity show promise in increasing number and/or function of these  
360 CPCs [45–47], potentially through promoting bone marrow production of progenitor  
361 cell subsets (although the origin of EPCs has been a topic of debate recently [48]) or  
362 via reducing inflammatory or pro-apoptotic stimuli in the circulation [49], thus  
363 enhancing survival of these cells in our body. Our data support the use of physical  
364 activity to promote or maintain CD34<sup>+</sup> CPC number in humans. High levels of self-  
365 reported physical activity were associated with reduced risk of all-cause mortality  
366 (**Supplementary Tables 3 and 4**), and they were associated with a higher number of  
367 CD34<sup>+</sup> CPCs, which were also associated with mortality, but only in individuals with  
368 CVD, and not in our CVD-free group.

369

370 Together these findings suggest that the observed protection of increased CD34<sup>+</sup>  
371 CPCs on mortality in a diseased population is partly driven by the physical activity  
372 levels of individuals. These findings may be clinically relevant as they are supportive  
373 of exercise-based cardiac rehabilitation and suggest an area for future interventions.

374 Whilst both acute aerobic and resistance exercise can promote progenitor cell release  
375 and improve pro-angiogenic function, long-term resistance exercise training studies  
376 are lacking and thus warranted, specifically in a CVD cohort.

377

#### 378 **4.1 Limitations**

379 The participants in this study self-reported physical activity levels, and thus, to  
380 confirm our findings, studies that include accelerometer-derived physical activity  
381 levels are required. This will allow researchers to more accurately assess the influence  
382 of light, moderate and strenuous activity, as well as inactivity, on measures of  
383 vascular repair and regeneration, key to maintenance of CV health. Additionally,  
384 repeated longitudinal measures of physical activity, CPCs and other clinical markers  
385 would provide more robust evidence for the relationship between physical activity  
386 and these markers of vascular repair. It must be noted that these findings are  
387 associative, and do not necessarily imply causality, however, there are several studies  
388 demonstrating the positive impact of exercise and physical activity on CPCs [45–47].  
389 Another consideration is the quantification of rare cells by flow cytometry. CD34<sup>+</sup>  
390 CPCs are typically between 0.001 and 0.01% of total circulating mononuclear cells  
391 [50], meaning accurate quantification can be problematic [51]. This limitation is  
392 compounded when investigating subpopulations, including CD34<sup>+</sup>KDR<sup>+</sup> CPCs, which  
393 are even fewer in number, therefore the predictive strength of these cells is reduced,  
394 despite evidence showing their positive impact on the vasculature.

395

#### 396 **4.2 Conclusions**

397 Our study demonstrated that CD34<sup>+</sup> and CD34<sup>+</sup>CD133<sup>+</sup> CPCs are predictive of  
398 mortality in a large cohort, but with more prognostic potential in individuals with

399 CVD. For the first time, we have provided data that shows physical activity is  
400 associated with significantly greater CD34<sup>+</sup> CPCs in a CVD population, with no  
401 relationship in a non-CVD population. Exercise and physical activity may promote  
402 vascular health and longevity in CVD patients via modulating CD34<sup>+</sup> CPC number.  
403

#### 404 **Declaration of competing interests**

405 The authors declare that they have no known competing financial interests or personal  
406 relationships that could have appeared to influence the work reported in this paper.  
407

#### 408 **Authors contributions**

409 M.R conceived and designed the research. D.M, M.R and J.D undertook statistical  
410 analysis of the data. M.R and D.M interpreted results of the experiments, prepared  
411 figures and drafted the manuscript; all authors edited and revised the manuscript; all  
412 authors approved the final version of the manuscript.  
413

#### 414 **Financial support**

415 This work was supported by Edinburgh Napier University's Research Excellence  
416 Grant (M.R). D.M. is supported by the European Union's INTERREG VA  
417 Programme, managed by the Special EU Programmes Body (SEUPB).  
418

419

420

#### 420 **Acknowledgements**

421 This manuscript was prepared manuscript was prepared using FRAMOFFSPRING  
422 Research Materials obtained from the NHLBI Biologic Specimen and Data

423 Repository Information Coordinating Center and does not necessarily reflect the  
 424 opinions or views of the FRAMCOHORT, FRAMOFFSPRING or the NHLBI.

425

426

427

## References

- 428 [1] S. Ananthaseshan, M.K. Grudzinska, K. Bojakowski, E. Kurzejamska, Z.  
 429 Gaciong, C. Söderberg-Nauclér, P. Religa, Locally Transplanted CD34+ Bone  
 430 Marrow-Derived Cells Contribute to Vascular Healing After Vascular Injury,  
 431 Transplantation Proceedings. 49 (2017) 1467–1476.  
 432 <https://doi.org/10.1016/j.transproceed.2017.01.081>.
- 433 [2] M. Herrmann, A. Binder, U. Menzel, S. Zeiter, M. Alini, S. Verrier,  
 434 CD34/CD133 enriched bone marrow progenitor cells promote  
 435 neovascularization of tissue engineered constructs in vivo, Stem Cell Research.  
 436 13 (2014) 465–477. <https://doi.org/10.1016/j.scr.2014.10.005>.
- 437 [3] T. Asahara, T. Murohara, A. Sullivan, M. Silver, R. van der Zee, T. Li, B.  
 438 Witzenbichler, G. Schatteman, J.M. Isner, Isolation of putative progenitor  
 439 endothelial cells for angiogenesis, Science. 275 (1997) 964–967.  
 440 <https://doi.org/10.1126/science.275.5302.964>.
- 441 [4] L. Bruyndonckx, V.Y. Hoymans, G. Frederix, A. de Guchtenaere, H. Franckx,  
 442 D.K. Vissers, C.J. Vrints, J. Ramet, V.M. Conraads, Endothelial progenitor  
 443 cells and endothelial microparticles are independent predictors of endothelial  
 444 function, Journal of Pediatrics. 165 (2014) 300–305.  
 445 <https://doi.org/10.1016/j.jpeds.2014.04.015>.
- 446 [5] N. Werner, S. Wassmann, P. Ahlers, T. Schiegl, S. Kosiol, A. Link, K.  
 447 Walenta, G. Nickenig, Endothelial progenitor cells correlate with endothelial  
 448 function in patients with coronary artery disease, Basic Research in  
 449 Cardiology. 102 (2007) 565–571. <https://doi.org/10.1007/s00395-007-0680-1>.
- 450 [6] E.M. van Craenenbroeck, G. Frederix, N. Pattyn, P. Beckers, A.H. van  
 451 Craenenbroeck, A. Gevaert, N. Possemiers, V. Cornelissen, K. Goetschalckx,  
 452 C.J. Vrints, L. Vanhees, V.Y. Hoymans, Effects of aerobic interval training and  
 453 continuous training on cellular markers of endothelial integrity in coronary  
 454 artery disease: A SAINTEX-CAD substudy, American Journal of Physiology -  
 455 Heart and Circulatory Physiology. 309 (2015) H1876–H1882.  
 456 <https://doi.org/10.1152/ajpheart.00341.2015>.
- 457 [7] L.E. Sidney, M.J. Branch, S.E. Dunphy, H.S. Dua, A. Hopkinson, Concise  
 458 review: Evidence for CD34 as a common marker for diverse progenitors, Stem  
 459 Cells. 32 (2014) 1380–1389. <https://doi.org/10.1002/stem.1661>.
- 460 [8] M. Hristov, W. Erl, P.C. Weber, Endothelial progenitor cells: Mobilization,  
 461 differentiation, and homing, Arteriosclerosis, Thrombosis, and Vascular  
 462 Biology. 23 (2003) 1185–1189.  
 463 <https://doi.org/10.1161/01.ATV.0000073832.49290.B5>.
- 464 [9] J.M. Hill, G. Zalos, J.P.J. Halcox, W.H. Schenke, M.A. Waclawiw, A.A.  
 465 Quyyumi, T. Finkel, Circulating Endothelial Progenitor Cells, Vascular  
 466 Function, and Cardiovascular Risk, New England Journal of Medicine. 348  
 467 (2003) 593–600. <https://doi.org/10.1056/NEJMoa022287>.

- 468 [10] C. Schmidt-Lucke, L. Rössig, S. Fichtlscherer, M. Vasa, M. Britten, U.  
 469 Kämper, S. Dimmeler, A.M. Zeiher, Reduced number of circulating endothelial  
 470 progenitor cells predicts future cardiovascular events: Proof of concept for the  
 471 clinical importance of endogenous vascular repair, *Circulation*. 111 (2005)  
 472 2981–2987. <https://doi.org/10.1161/CIRCULATIONAHA.104.504340>.
- 473 [11] A. Mehta, A.S. Tahhan, C. Liu, D.S. Dhindsa, A. Nayak, A. Hooda, K.  
 474 Moazzami, S.J. Islam, S.C. Rogers, Z. Almuwaqqat, A. Mokhtari, I. Hesaroieh,  
 475 Y.A. Ko, E.K. Waller, A.A. Quyyumi, Circulating Progenitor Cells in Patients  
 476 With Coronary Artery Disease and Renal Insufficiency, *JACC: Basic to*  
 477 *Translational Science*. 5 (2020) 770–782.  
 478 <https://doi.org/10.1016/j.jacbts.2020.06.006>.
- 479 [12] G.P. Fadini, A. Coracina, I. Baesso, C. Agostini, A. Tiengo, A. Avogaro, S.V.  
 480 de Kreutzenberg, Peripheral blood CD34+KDR+ endothelial progenitor cells  
 481 are determinants of subclinical atherosclerosis in a middle-aged general  
 482 population, *Stroke*. 37 (2006) 2277–2282.  
 483 <https://doi.org/10.1161/01.STR.0000236064.19293.79>.
- 484 [13] A. Samman Tahhan, M. Hammadah, P.B. Sandesara, S.S. Hayek, A.P.  
 485 Kalogeropoulos, A. Alkholder, H. Mohamed Kelli, M. Topel, N. Ghasemzadeh,  
 486 K. Chivukula, Y.A. Ko, H. Aida, I. Hesaroieh, E. Mahar, J.H. Kim, P. Wilson,  
 487 L. Shaw, V. Vaccarino, E.K. Waller, A.A. Quyyumi, Progenitor Cells and  
 488 Clinical Outcomes in Patients with Heart Failure, *Circulation: Heart Failure*. 10  
 489 (2017). <https://doi.org/10.1161/CIRCHEARTFAILURE.117.004106>.
- 490 [14] A.S. Tahhan, M. Hammadah, M. Raad, Z. Almuwaqqat, A. Alkholder, P.B.  
 491 Sandesara, H. Mohamed-Kelli, S.S. Hayek, J.H. Kim, W.T. O’Neal, M.L.  
 492 Topel, A.J. Grant, N. Sabbak, R.E. Heintz, M.M. Gafeer, M. Obideen, B.  
 493 Kaseer, N. Abdelhadi, Y.A. Ko, C. Liu, I. Hesaroieh, E.A. Mahar, V.  
 494 Vaccarino, E.K. Waller, A.A. Quyyumi, Progenitor Cells and Clinical  
 495 Outcomes in Patients with Acute Coronary Syndromes, *Circulation Research*.  
 496 122 (2018) 1565–1575. <https://doi.org/10.1161/CIRCRESAHA.118.312821>.
- 497 [15] A. Burke, G.A. FitzGerald, Oxidative stress and smoking-induced vascular  
 498 injury, *Progress in Cardiovascular Diseases*. 46 (2003) 79–90.  
 499 [https://doi.org/10.1016/S0033-0620\(03\)00076-8](https://doi.org/10.1016/S0033-0620(03)00076-8).
- 500 [16] U. Laufs, S. Wassmann, T. Czech, T. Münzel, M. Eisenhauer, M. Böhm, G.  
 501 Nickenig, Physical inactivity increases oxidative stress, endothelial  
 502 dysfunction, and atherosclerosis, *Arteriosclerosis, Thrombosis, and Vascular*  
 503 *Biology*. 25 (2005) 809–814.  
 504 <https://doi.org/10.1161/01.ATV.0000158311.24443.af>.
- 505 [17] O.J. MacEaney, E.J. Kushner, G.P. van Guilder, J.J. Greiner, B.L. Stauffer,  
 506 C.A. DeSouza, Endothelial progenitor cell number and colony-forming  
 507 capacity in overweight and obese adults, *International Journal of Obesity*. 33  
 508 (2009) 219–225. <https://doi.org/10.1038/ijo.2008.262>.
- 509 [18] D.R. Seals, E.E. Nagy, K.L. Moreau, Aerobic exercise training and vascular  
 510 function with ageing in healthy men and women, *Journal of Physiology*. 597  
 511 (2019) 4901–4914. <https://doi.org/10.1113/JP277764>.
- 512 [19] A. Campbell, F. Grace, L. Ritchie, A. Beaumont, N. Sculthorpe, Long-term  
 513 aerobic exercise improves vascular function into old age: A systematic review,  
 514 meta-analysis and meta regression of observational and interventional studies,  
 515 *Frontiers in Physiology*. 10 (2019). <https://doi.org/10.3389/fphys.2019.00031>.
- 516 [20] M.D. Ross, A.L. Wekesa, J.P. Phelan, M. Harrison, Resistance exercise  
 517 increases endothelial progenitor cells and angiogenic factors, *Medicine and*



- 518 Science in Sports and Exercise. 46 (2014) 16–23.  
 519 <https://doi.org/10.1249/MSS.0b013e3182a142da>.
- 520 [21] N.H. Agha, F.L. Baker, H.E. Kunz, R. Graff, R. Azadan, C. Dolan, M.S.  
 521 Laughlin, C. Hosing, M.M. Markofski, R.A. Bond, C.M. Bollard, R.J.  
 522 Simpson, Vigorous exercise mobilizes CD34+ hematopoietic stem cells to  
 523 peripheral blood via the  $\beta$  2 -adrenergic receptor, *Brain, Behavior, and*  
 524 *Immunity*. 68 (2018) 66–75. <https://doi.org/10.1016/j.bbi.2017.10.001>.
- 525 [22] M. Sandri, E.B. Beck, V. Adams, S. Gielen, K. Lenk, R. Höllriegel, N.  
 526 Mangner, A. Linke, S. Erbs, S. Möbius-Winkler, D. Scheinert, R. Hambrecht,  
 527 G. Schuler, Maximal exercise, limb ischemia, and endothelial progenitor cells,  
 528 *European Journal of Preventive Cardiology*. 18 (2011) 55–64.  
 529 <https://doi.org/10.1097/HJR.0b013e32833ba654>.
- 530 [23] E.M. van Craenenbroeck, L. Bruyndonckx, C. van Berckelaer, V.Y. Hoymans,  
 531 C.J. Vrints, V.M. Conraads, The effect of acute exercise on endothelial  
 532 progenitor cells is attenuated in chronic heart failure, *European Journal of*  
 533 *Applied Physiology*. 111 (2011) 2375–2379. [https://doi.org/10.1007/s00421-](https://doi.org/10.1007/s00421-011-1843-1)  
 534 [011-1843-1](https://doi.org/10.1007/s00421-011-1843-1).
- 535 [24] S. Steiner, A. Niessner, S. Ziegler, B. Richter, D. Seidinger, J. Pleiner, M.  
 536 Penka, M. Wolzt, K. Huber, J. Wojta, E. Minar, C.W. Kopp, Endurance  
 537 training increases the number of endothelial progenitor cells in patients with  
 538 cardiovascular risk and coronary artery disease, *Atherosclerosis*. 181 (2005)  
 539 305–310. <https://doi.org/10.1016/j.atherosclerosis.2005.01.006>.
- 540 [25] M. Brehm, F. Picard, P. Ebner, G. Turan, E. Bölke, M. Köstering, P. Schüller,  
 541 T. Fleissner, D. Ilousis, K. Augusta, M. Peiper, C. Schannwell, B.E. Strauer,  
 542 Effects of exercise training on mobilization and functional activity of blood-  
 543 derived progenitor cells in patients with acute myocardial infarction, *European*  
 544 *Journal of Medical Research*. 14 (2009) 393–405. [https://doi.org/10.1186/2047-](https://doi.org/10.1186/2047-783x-14-9-393)  
 545 [783x-14-9-393](https://doi.org/10.1186/2047-783x-14-9-393).
- 546 [26] W.B. Kannel, T. Gordon, Evaluation of cardiovascular risk in the elderly: The  
 547 Framingham study, *Bulletin of the New York Academy of Medicine: Journal*  
 548 *of Urban Health*. 54 (1978) 573–591.  
 549 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1807497/> (accessed May 3,  
 550 2021).
- 551 [27] W. Kannel, M. Feinleib, P. McNamara, R. Garrison, W. Castelli, An  
 552 investigation of coronary heart disease in families, *American Journal of*  
 553 *Epidemiology*. 110 (1979) 281–290.  
 554 <https://doi.org/10.1093/oxfordjournals.aje.a112813>.
- 555 [28] S. Cheng, N. Wang, M.G. Larson, J.N. Palmisano, G.F. Mitchell, E.J.  
 556 Benjamin, R.S. Vasan, D. Levy, E.L. McCabe, J.A. Vita, T.J. Wang, S.Y.  
 557 Shaw, K.S. Cohen, N.M. Hamburg, Circulating angiogenic cell populations,  
 558 vascular function, and arterial stiffness, *Atherosclerosis*. 220 (2012) 145–150.  
 559 <https://doi.org/10.1016/j.atherosclerosis.2011.10.015>.
- 560 [29] W.B. Kannel, P. Sorlie, Some Health Benefits of Physical Activity: The  
 561 Framingham Study, *Archives of Internal Medicine*. 139 (1979) 857–861.  
 562 <https://doi.org/10.1001/archinte.1979.03630450011006>.
- 563 [30] P.W.F. Wilson, R.S. Paffenbarger, J.N. Morris, R.J. Havlik, Assessment  
 564 methods for physical activity and physical fitness in population studies: Report  
 565 of a NHLBI workshop, *American Heart Journal*. 111 (1986) 1177–1192.  
 566 [https://doi.org/10.1016/0002-8703\(86\)90022-0](https://doi.org/10.1016/0002-8703(86)90022-0).

- 567 [31] R.S. Patel, Q. Li, N. Ghasemzadeh, D.J. Eapen, L.D. Moss, A.U. Janjua, P.  
568 Manocha, H. al Kassem, E. Veledar, H. Samady, W.R. Taylor, A.M. Zafari, L.  
569 Sperling, V. Vaccarino, E.K. Waller, A.A. Quyyumi, Circulating CD34+  
570 progenitor cells and risk of mortality in a population with coronary artery  
571 disease, *Circulation Research*. 116 (2015) 289–297.  
572 <https://doi.org/10.1161/CIRCRESAHA.116.304187>.
- 573 [32] C.G. Egan, C. Fondelli, E. Pierantozzi, G. Tripepi, F. Dotta, V. Sorrentino,  
574 Putative endothelial progenitor cells predict long-term mortality in type-2  
575 diabetes, *Endocrine*. 62 (2018) 263–266. [https://doi.org/10.1007/s12020-018-](https://doi.org/10.1007/s12020-018-1695-0)  
576 [1695-0](https://doi.org/10.1007/s12020-018-1695-0).
- 577 [33] N. Werner, S. Kosiol, T. Schiegl, P. Ahlers, K. Walenta, A. Link, M. Böhm, G.  
578 Nickenig, Circulating Endothelial Progenitor Cells and Cardiovascular  
579 Outcomes, *New England Journal of Medicine*. 353 (2005) 999–1007.  
580 <https://doi.org/10.1056/nejmoa043814>.
- 581 [34] M. Peichev, A.J. Naiyer, D. Pereira, Z. Zhu, W.J. Lane, M. Williams, M.C. Oz,  
582 D.J. Hicklin, L. Witte, M.A.S. Moore, S. Rafii, Expression of VEGFR-2 and  
583 AC133 by circulating human CD34+ cells identifies a population of functional  
584 endothelial precursors, *Blood*. 95 (2000) 952–958.  
585 [https://doi.org/10.1182/blood.v95.3.952.003k27\\_952\\_958](https://doi.org/10.1182/blood.v95.3.952.003k27_952_958).
- 586 [35] E.B. Friedrich, K. Walenta, J. Scharlau, G. Nickenig, N. Werner, CD34-  
587 /CD133+/VEGFR-2+ endothelial progenitor cell subpopulation with potent  
588 vasoregenerative capacities, *Circulation Research*. 98 (2006).  
589 <https://doi.org/10.1161/01.RES.0000205765.28940.93>.
- 590 [36] J. Hur, C.H. Yoon, H.S. Kim, J.H. Choi, H.J. Kang, K.K. Hwang, B.H. Oh,  
591 M.M. Lee, Y.B. Park, Characterization of Two Types of Endothelial Progenitor  
592 Cells and Their Different Contributions to Neovasclogenesis, *Arteriosclerosis,*  
593 *Thrombosis, and Vascular Biology*. 24 (2004) 288–293.  
594 <https://doi.org/10.1161/01.ATV.0000114236.77009.06>.
- 595 [37] B.A. Nasser, W. Ebell, M. Dandel, M. Kukucka, R. Gebker, A. Doltra, C.  
596 Knosalla, Y.H. Choi, R. Hetzer, C. Stamm, Autologous CD133+ bone marrow  
597 cells and bypass grafting for regeneration of ischaemic myocardium: The  
598 Cardio133 trial, *European Heart Journal*. 35 (2014) 1263–1274.  
599 <https://doi.org/10.1093/eurheartj/ehu007>.
- 600 [38] A.A. Quyyumi, A. Vasquez, D.J. Kereiakes, M. Klapholz, G.L. Schaer, A.  
601 Abdel-Latif, S. Frohwein, T.D. Henry, R.A. Schatz, N. Dib, C. Toma, C.J.  
602 Davidson, G.W. Barsness, D.M. Shavelle, M. Cohen, J. Poole, T. Moss, P.  
603 Hyde, A.M. Kanakaraj, V. Druker, A. Chung, C. Junge, R.A. Preti, R.L. Smith,  
604 D.J. Mazzo, A. Pecora, D.W. Losordo, PreSERVE-AMI: A Randomized,  
605 Double-Blind, Placebo-Controlled Clinical Trial of Intracoronary  
606 Administration of Autologous CD34+ Cells in Patients with Left Ventricular  
607 Dysfunction Post STEMI, *Circulation Research*. 120 (2017) 324–331.  
608 <https://doi.org/10.1161/CIRCRESAHA.115.308165>.
- 609 [39] M.C. Barsotti, T. Santoni, M.E.L. Picoi, N. Mancini, F. Massaro, C.  
610 Grigoratos, U. Bortolotti, P. Collecchi, M. Menicagli, C. Scatena, F. Felice, G.  
611 Bevilacqua, A.G. Naccarato, R. di Stefano, A. Balbarini, Endothelial  
612 progenitor cell homing in human myocardium in patients with coronary artery  
613 disease, *International Journal of Cardiology*. 172 (2014) 516–517.  
614 <https://doi.org/10.1016/j.ijcard.2014.01.042>.
- 615 [40] Y.F. Liao, Y. Feng, L.L. Chen, T.S. Zeng, F. Yu, L.J. Hu, Coronary heart  
616 disease risk equivalence in diabetes and arterial diseases characterized by



- 617 endothelial function and endothelial progenitor cell, *Journal of Diabetes and Its*  
 618 *Complications*. 28 (2014) 214–218.  
 619 <https://doi.org/10.1016/j.jdiacomp.2013.09.009>.
- [41] J.P. Nederveen, J. Baker, G. Ibrahim, V. Ivankovic, M.E. Percival, G. Parise,  
 620 Hematopoietic Stem and Progenitor Cell (HSPC) Mobilization Responses to  
 621 Different Exercise Intensities in Young and Older Adults, *Journal of Science in*  
 622 *Sport and Exercise*. 2 (2020) 47–58. [https://doi.org/10.1007/s42978-019-](https://doi.org/10.1007/s42978-019-00050-4)  
 623 [00050-4](https://doi.org/10.1007/s42978-019-00050-4).  
 624
- [42] E.M.F. van Craenenbroeck, C.J. Vrints, S.E. Haine, K. Vermeulen, I.  
 625 Goovaerts, V.F.I. van Tendeloo, V.Y. Hoymans, V.M.A. Conraads, A maximal  
 626 exercise bout increases the number of circulating CD34+/KDR+ endothelial  
 627 progenitor cells in healthy subjects. Relation with lipid profile, *Journal of*  
 628 *Applied Physiology*. 104 (2008) 1006–1013.  
 629 <https://doi.org/10.1152/jappphysiol.01210.2007>.
- [43] V. Adams, K. Lenk, A. Linke, D. Lenz, S. Erbs, M. Sandri, A. Tarnok, S.  
 630 Gielen, F. Emmrich, G. Schuler, R. Hambrecht, Increase of Circulating  
 631 Endothelial Progenitor Cells in Patients with Coronary Artery Disease after  
 632 Exercise-Induced Ischemia, *Arteriosclerosis, Thrombosis, and Vascular*  
 633 *Biology*. 24 (2004) 684–690.  
 634 <https://doi.org/10.1161/01.ATV.0000124104.23702.a0>.
- [44] K. Moazzami, B.B. Lima, M. Hammadah, R. Ramadan, I. al Mheid, J.H. Kim,  
 635 A. Alkhoder, M. Obideen, O. Levantsevych, A. Shah, C. Liu, J.D. Bremner, M.  
 636 Kutner, Y. v. Sun, E.K. Waller, I.G. Hesaroieh, P. Raggi, V. Vaccarino, A.A.  
 637 Quyyumi, Association between Change in Circulating Progenitor Cells during  
 638 Exercise Stress and Risk of Adverse Cardiovascular Events in Patients with  
 639 Coronary Artery Disease, *JAMA Cardiology*. 5 (2020) 147–155.  
 640 <https://doi.org/10.1001/jamacardio.2019.4528>.
- [45] S.L. Cavalcante, S. Lopes, L. Bohn, I. Caverro-Redondo, C. Álvarez-Bueno, S.  
 641 Viamonte, M. Santos, J. Oliveira, F. Ribeiro, Effects of exercise on endothelial  
 642 progenitor cells in patients with cardiovascular disease: A systematic review  
 643 and meta-analysis of randomized controlled trials, *Revista Portuguesa de*  
 644 *Cardiologia*. 38 (2019) 817–827. <https://doi.org/10.1016/j.repc.2019.02.016>.
- [46] L. Bruyndonckx, V.Y. Hoymans, A. de Guchtenaere, M. van Helvoirt, E.M.  
 645 van Craenenbroeck, G. Frederix, K. Lemmens, D.K. Vissers, C.J. Vrints, J.  
 646 Ramet, V.M. Conraads, Diet, exercise, and endothelial function in obese  
 647 adolescents, *Pediatrics*. 135 (2015) e653–e661.  
 648 <https://doi.org/10.1542/peds.2014-1577>.
- [47] Y. Guo, R.A. Ledesma, R. Peng, Q. Liu, D. Xu, The Beneficial Effects of  
 649 Cardiac Rehabilitation on the Function and Levels of Endothelial Progenitor  
 650 Cells, *Heart Lung and Circulation*. 26 (2017) 10–17.  
 651 <https://doi.org/10.1016/j.hlc.2016.06.1210>.
- [48] T. Fujisawa, O. Tura-Ceide, A. Hunter, A. Mitchell, A. Vesey, C. Medine, S.  
 652 Gallogly, P.W.F. Hadoke, C. Keith, A. Sproul, H. Roddie, G. McQuaker, I.  
 653 Wilmut, N.L. Mills, M. Brittan, Endothelial Progenitor Cells Do Not Originate  
 654 From the Bone Marrow, *Circulation*. 140 (2019) 1524–1526.  
 655 <https://doi.org/10.1161/CIRCULATIONAHA.119.042351>.
- [49] F. Cesari, R. Marcucci, A.M. Gori, C. Burgisser, S. Francini, F. Sofi, G.F.  
 656 Gensini, R. Abbate, F. Fattiroli, Impact of a cardiac rehabilitation program and  
 657 inflammatory state on endothelial progenitor cells in acute coronary syndrome  
 658

- 666 patients, *International Journal of Cardiology*. 167 (2013) 1854–1859.  
667 <https://doi.org/10.1016/j.ijcard.2012.04.157>.
- 668 [50] J. Case, L.E. Mead, W.K. Bessler, D. Prater, H.A. White, M.R. Saadatzadeh,  
669 J.R. Bhavsar, M.C. Yoder, L.S. Haneline, D.A. Ingram, Human  
670 CD34+AC133+VEGFR-2+ cells are not endothelial progenitor cells but  
671 distinct, primitive hematopoietic progenitors, *Experimental Hematology*. 35  
672 (2007) 1109–1118. <https://doi.org/10.1016/j.exphem.2007.04.002>.
- 673 [51] E.M. van Craenenbroeck, A.H. van Craenenbroeck, S. van Ierssel, L.  
674 Bruyndonckx, V.Y. Hoymans, C.J. Vrints, V.M. Conraads, Quantification of  
675 circulating CD34+/KDR+/CD45 dim endothelial progenitor cells: Analytical  
676 considerations, *International Journal of Cardiology*. 167 (2013) 1688–1695.  
677 <https://doi.org/10.1016/j.ijcard.2012.10.047>.  
678
- 679
- 680

681 **Figure legends**

682

683 **Figure 1.** Flow chart of the Framingham Offspring Cohort and the participants included  
684 in this study.

685 MI- myocardial infarction, HF- heart failure, IC- intermittent claudication.

686

687 **Figure 2.** Kaplan Meier survival curve's for the relationship between CPC tertile group  
688 (CD34<sup>+</sup> n=1751, CD34<sup>+</sup>CD133<sup>+</sup> n=1630, CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup> n=1751, ECFC  
689 n=1649) and all-cause mortality (A-D) and cardiovascular mortality (E-H) (Tertile 1 =  
690 Low count, Tertile 2 = Moderate count, Tertile 3 = High count).

691 Statistical significance was set at  $p < 0.05$  derived from Cox proportional hazard  
692 regressions.

693

694

695

696

697

698

699

700

701

702

703

704

705

706

707

708

709

710

711

712

713

714

715

716 **Tables**717 **Table 1.** Participant characteristics

	All (n=1751)	CVD-free (n=1467, 84%)	CVD (n=284, 16%)	<i>p</i> -value
Age (years)	66 ± 9 [40-92]	65 ± 9 [40-90]	72 ± 9 [51-92]	<0.001
Female (n, %)	940, 53.7%	822, 56%	118, 41.5%	-
BMI (kg·m <sup>2</sup> )	28.4 ± 5.3 [13.8-54.2]	28.2 ± 5.3 [13.8-54.2]	29.2 ± 5.2 [18.4-45.1]	0.007
Systolic blood Pressure (mmHg)	129 ± 18 [76-204]	129 ± 18 [76-204]	131 ± 19 [90-198]	0.070
Diastolic blood Pressure (mmHg)	74 ± 11 [34-122]	75 ± 10 [34-122]	70 ± 11 [40-108]	<0.001***
Fasting glucose (mg/dL)	107.0 ± 24.1 [36-327]	106 ± 23 [58-327]	113 ± 28 [36-292]	<0.001***
Total cholesterol (mg/dL)	186.0 ± 37.5 [71-322]	190 ± 35.6 [96-322]	164 ± 39.4 [71-289]	<0.001***
HDL-cholesterol (mg/dL)	57.4 ± 18.2 [21-147]	58.8 ± 18.3 [21-152]	50.3 ± 15.8 [23-130]	<0.001***
Triglycerides (mg/dL)	118.0 ± 70.6 [30-976]	115 ± 67.2 [30-976]	133 ± 84.4 [40-583]	<0.001
Smokers (n, %)	206, 11.8%	170, 12%	36, 13%	
Hypertensive (n, %)	989, 56.5%	831, 57%	158, 56%	-
CD34+ CPCs (% MNCs)	0.0873 ± 0.0492 [0.0111-0.490]	0.0875 ± 0.0483 [0.0110-0.490]	0.0860 ± 0.0538 [0.0160-0.370]	0.201
CD34+CD133+ CPCs (% MNCs)	0.0402 ± 0.0366 [0.0020-0.6090]	0.0407 ± 0.0380 [0.0020-0.6090]	0.0375 ± 0.0286 [0.0040-0.2420]	0.290
	(n=1630, 55% Female)	(n=1356, 58% Female)	(n=274, 41% Female)	

CD34 <sup>+</sup> CD133 <sup>+</sup> KDR <sup>+</sup>	0.0040 ± 0.0037	0.0040 ± 0.0037	0.0041 ± 0.0039	0.648
(% MNCs)	[0.0001-0.0470]	[0.0001-0.0470]	[0.0002-0.0261]	
ECFC (number of colonies)	43 ± 31 [0-196]	43 ± 31 [0-196]	42 ± 32 [0-178]	0.470
	(n=1649, 53% female)	(n=1387, 56% female)	(n=262, 41% female)	

718 Data are mean ± SD [range]. CD34<sup>+</sup>CD133<sup>+</sup> (in 1630 participants).

719 ECFC- endothelial colony forming cells (in 1649 participants). \*  $p < 0.05$ , \*\*\*  $p$

720  $< 0.001$ , independent samples T-test.

721

722 **Table 2.** CPC Counts and mortality risk

CPC subset	Outcome	Model	No. of events/ No. at risk	HR	HR 95% CI	<i>p</i> value
<b>CD34<sup>+</sup> CPCs</b>	All-cause mortality	Unadjusted	326/1751	0.64	0.52 - 0.79	<0.001***
		Adjusted <sup>a</sup>	326/1751	0.79	0.64 - 0.98	0.036*
	CVD mortality	Unadjusted	71/1751	0.54	0.35 - 0.85	0.008**
		Adjusted <sup>a</sup>	71/1751	0.64	0.41 - 1.01	0.055
<b>CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup> EPCs</b>	All-cause mortality	Unadjusted	326/1751	0.88	0.76 - 1.01	0.066
		Adjusted <sup>a</sup>	326/1751	0.93	0.80 - 1.07	0.300
	CVD mortality	Unadjusted	71/1751	1.02	0.76 - 1.38	0.888
		Adjusted <sup>a</sup>	71/1751	1.09	0.81 - 1.47	0.579
<b>CD34<sup>+</sup>CD133<sup>+</sup> CPCs</b>	All-cause mortality	Unadjusted	303/1630	0.76	0.65 - 0.90	0.001**
		Adjusted <sup>a</sup>	303/1630	0.86	0.72 - 1.01	0.07
	CVD mortality	Unadjusted	65/1630	0.61	0.43 - 0.87	0.006**
		Adjusted <sup>a</sup>	65/1630	0.63	0.44 - 0.91	0.013*
<b>ECFC</b>	All-cause mortality	Unadjusted	321/1649	0.96	0.92 - 1.04	0.11
		Adjusted <sup>a</sup>	321/1649	0.98	0.94 - 1.03	0.393
	CVD mortality	Unadjusted	71/1649	0.97	0.88 - 1.07	0.505
		Adjusted <sup>a</sup>	71/1649	0.99	0.90 - 1.10	0.893

723 HR – hazard ratio, CI – confidence intervals.

724 <sup>a</sup>Model adjusted for age, sex, BMI, PAI, smoking status, diabetes status, hypertension  
725 and previous CVD diagnosis.726 \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$ , \*  $p < 0.05$  derived from Cox proportional hazard regressions

727 **Table 3.** CPC Counts and risk of death for all participants split by CVD diagnosis at exam 8

CPC Subset	Outcome	Model	CVD free at exam 8				CVD diagnosis by exam 8			
			No. of events/ No. at risk	HR	HR 95% CI	P value	No. of events/ No. at risk	HR	HR 95% CI	P value
<b>CD34<sup>+</sup> CPCs</b>	All-cause mortality	Unadjusted	211/1467	0.70	0.54 - 0.91	0.008**	115/284	0.57	0.41 - 0.81	0.002**
		Adjusted <sup>a</sup>	211/1467	0.89	0.67 - 1.18	0.424	115/284	0.68	0.48 - 0.97	0.032*
	CVD mortality	Unadjusted	34/1467	0.65	0.34 - 1.26	0.203	37/284	0.49	0.26 - 0.91	0.023*
		Adjusted <sup>a</sup>	34/1467	0.84	0.41 - 1.70	0.619	37/284	0.53	0.28 - 0.99	0.048*
<b>CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup> EPCs</b>	All-cause mortality	Unadjusted	211/1467	0.90	0.76 - 1.08	0.258	115/284	0.85	0.68 - 1.06	0.149
		Adjusted <sup>a</sup>	211/1467	0.92	0.76 - 1.11	0.385	115/284	0.94	0.75 - 1.18	0.586
	CVD mortality	Unadjusted	34/1467	0.86	0.56 - 1.33	0.492	37/284	1.17	0.79 - 1.72	0.442
		Adjusted <sup>a</sup>	34/1467	0.80	0.49 - 1.28	0.350	37/284	1.28	0.86 - 1.90	0.229
<b>CD34<sup>+</sup>CD133<sup>+</sup> CPCs</b>	All-cause mortality	Unadjusted	196/1364	0.85	0.69 - 1.04	0.11	107/266	0.61	0.46 - 0.82	0.001**
		Adjusted <sup>a</sup>	196/1364	0.99	0.80 - 1.23	0.943	107/266	0.64	0.48 - 0.86	0.003**
	CVD mortality	Unadjusted	31/1364	0.78	0.47 - 1.30	0.344	34/266	0.45	0.27 - 0.76	0.003**
		Adjusted <sup>a</sup>	31/1364	0.86	0.50 - 1.49	0.588	34/266	0.42	0.24 - 0.72	0.002**
<b>ECFC</b>	All-cause mortality	Unadjusted	205/1391	0.96	0.91 - 1.02	0.179	116/262	0.99	0.92 - 1.07	0.782
		Adjusted <sup>a</sup>	205/1391	0.97	0.92 - 1.02	0.261	116/262	1.01	0.93 - 1.09	0.828
	CVD mortality	Unadjusted	34/1391	1.03	0.90 - 1.19	0.655	37/262	0.93	0.82 - 1.07	0.317
		Adjusted <sup>a</sup>	34/1391	1.03	0.90 - 1.19	0.633	37/262	0.95	0.82 - 1.09	0.437

728 HR- hazard ratio, CI- confidence intervals.

729 <sup>a</sup>Model adjusted for age, sex, BMI, PAI, smoking status, diabetes status, hypertension. \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$ , \*  $p < 0.05$ 

730 derived from Cox proportional hazard regressions.

731 **Table 4.** Association between physical activity index and CPC Counts

CPC subset	Outcome	All	CVD-free	CVD
		$\beta$ , <i>T</i> -value, <i>p</i> -value	$\beta$ , <i>T</i> -value, <i>p</i> -value	$\beta$ , <i>T</i> -value, <i>p</i> -value
<b>CD34<sup>+</sup></b>	Unadjusted	0.023, 0.967, 0.334	-0.013, -0.486, 0.627	0.176, 3.009, 0.003**
	Adjusted <sup>a</sup>	0.008, 0.322, 0.748	-0.021, -0.813, 0.416	0.153, 2.461, 0.014*
<b>CD34<sup>+</sup>CD133<sup>+</sup></b>	Unadjusted	-0.014, -0.567, 0.571	-0.04, -1.495, 0.135	0.115, 1.893, 0.059
	Adjusted <sup>a</sup>	-0.022, -0.872, 0.383	-0.042, -1.574, 0.116	0.107, 1.616, 0.107
<b>CD34<sup>+</sup>CD133<sup>+</sup>KDR<sup>+</sup></b>	Unadjusted	0.022, 0.901, 0.368	0.002, 0.086, 0.932	0.108, 1.817, 0.07
	Adjusted <sup>a</sup>	0.019, 0.775, 0.439	0.005, 0.194, 0.846	0.080, 1.269, 0.205
<b>ECFC</b>	Unadjusted	0.005, 0.197, 0.844	-0.005, -0.201, 0.841	0.049, 0.785, 0.433
	Adjusted <sup>a</sup>	-0.006, -0.241, 0.809	-0.015, -0.535, 0.592	0.036, 0.537, 0.592

732 <sup>a</sup>Model adjusted for age, sex, BMI, smoking status, diabetes status and hypertension.

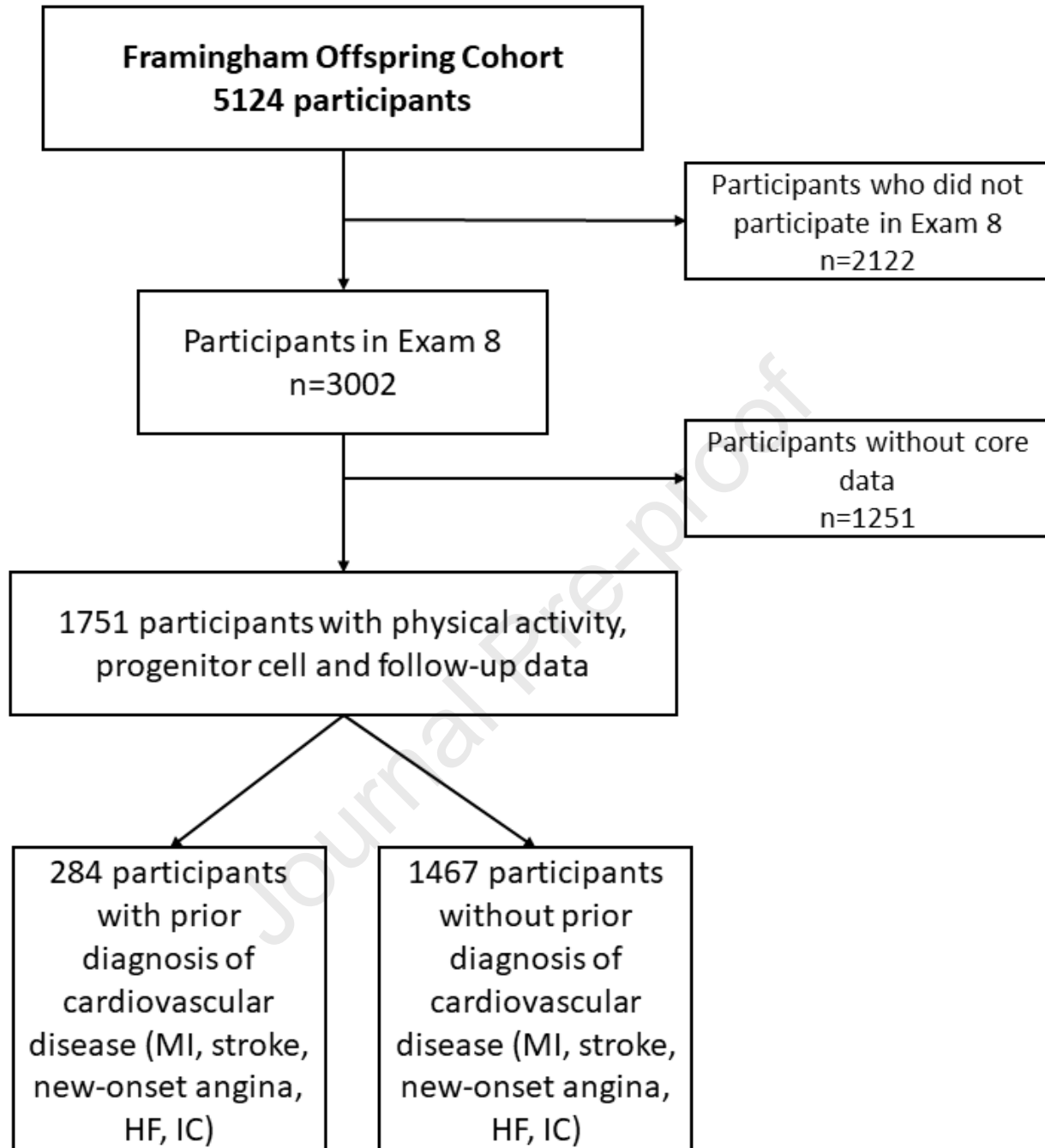
733 For “All” adjustment also includes previous CVD diagnosis

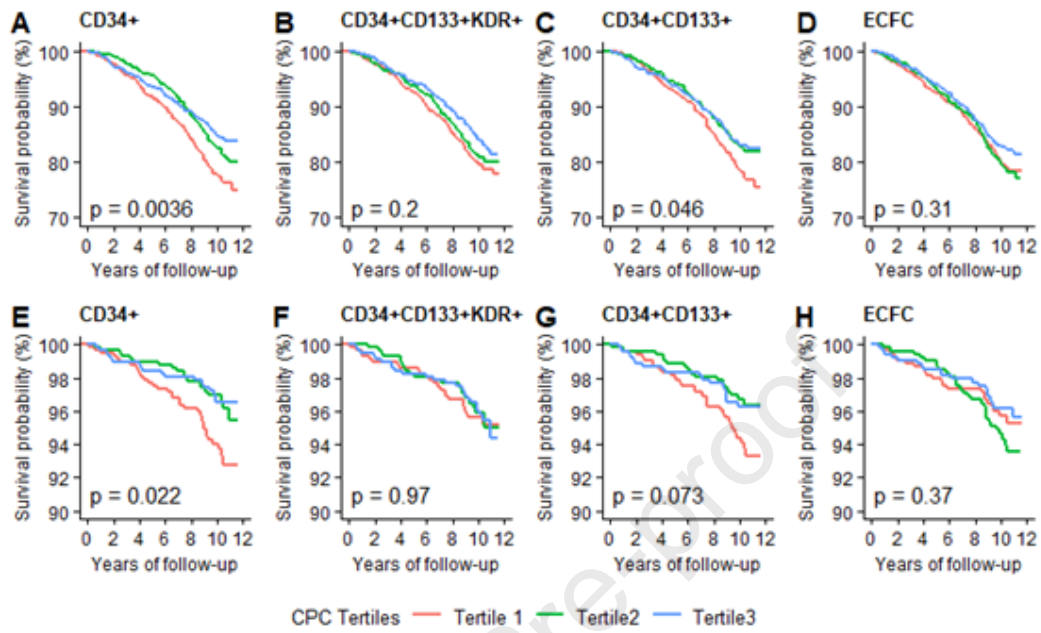
734 ECFC- endothelial colony forming cells.

735 \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$ , \*  $p < 0.05$  derived from Cox proportional hazard

736 regressions.







**CD34+ progenitors are predictive of mortality and are associated with physical activity  
in cardiovascular disease patients**

David Muggeridge<sup>ab</sup> ([D.Muggeridge@napier.ac.uk](mailto:D.Muggeridge@napier.ac.uk)), Jennifer Dodd<sup>a</sup> ([J.Dodd@napier.ac.uk](mailto:J.Dodd@napier.ac.uk)),

Mark D. Ross<sup>a</sup> ([M.Ross@napier.ac.uk](mailto:M.Ross@napier.ac.uk))

<sup>a</sup>School of Applied Sciences, Edinburgh Napier University, Edinburgh, United Kingdom

<sup>b</sup>Institute of Health Research & Innovation, Division of Biomedical Science, University of the Highlands and Islands, Inverness, UK

**Highlights:**

- Circulating CD34+ progenitor cells are biomarkers of endothelial regenerative capacity
- Lower levels of these cells are predictive of cardiovascular and all-cause mortality, specifically in those with underlying or pre-diagnosed cardiovascular disease
- Self-reported physical activity is positively associated with these CD34+ progenitor cells independent of other known risk factors, but only in individuals with pre-existing cardiovascular disease which may help to explain the role of physical activity in reducing future event risk

**Declaration of interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Journal Pre-proof