

# Ecological status and sources of anthropogenic contaminants in mangroves of the Wouri River Estuary (Cameroon)

Marco Fusi<sup>1,\*</sup>, Gian Maria Beone<sup>2</sup>, Nicoleta Alina Suci<sup>2</sup>, Angela Sacchi<sup>2</sup>, Marco Trevisan<sup>2</sup>, Ettore Capri<sup>2</sup>, Daniele Daffonchio<sup>1</sup>, Ndongo Din<sup>3</sup>, Farid Dahdouh-Guebas<sup>4,5,†</sup>, Stefano Cannicci<sup>6,7,†</sup>

## Affiliations

<sup>1</sup>King Abdullah University of Science and Technology (KAUST), Biological and Environmental Sciences & Engineering Division, Thuwal 23955-6900, Saudi Arabia;

<sup>2</sup>Institute of Agricultural and Environmental Chemistry, Università Cattolica del Sacro Cuore, Via Emilia Parmense 84, 29122 Piacenza, Italy.

<sup>3</sup>The University of Douala, Faculty of Science, Department of Botany, P.O. Box 8948 Douala Cameroon.

<sup>4</sup>Laboratory of Systems Ecology and Resource Management, Department of Organism Biology, Faculty of Sciences, Université Libre de Bruxelles – ULB, Avenue F.D. Roosevelt 50, CPI 264/1, B-1050 Brussels, Belgium.

<sup>5</sup>Laboratory of Plant Biology and Nature Management, Department of Biology, Faculty of Sciences and Bio-engineering Sciences, Vrije Universiteit Brussel – VUB, Pleinlaan 2, B-1050 Brussels, Belgium

<sup>6</sup>The Swire Institute of Marine Science and The School of Biological Sciences, The University of Hong Kong, Hong Kong

<sup>7</sup>Department of Biology, University of Florence, Via Madonna del Piano 6, Sesto Fiorentino, Italy.

† These authors equally contributed to the paper

## \*Corresponding author:

Email address: marco.fusi@kaust.edu.sa

## Abstract

Mangroves are critically threatened by human activities, despite the important ecosystem functions and services they provide. Mangroves in Cameroon represent no exception to the worldwide trend of mangrove destruction, especially around Douala, on the Wouri river estuary. In two sites around Douala, we assessed the presence of sterols, PAHs, PCBs, DEHP, DDT and its metabolite p-p'DDE and potentially toxic metals in sediment samples. As a proxy of ecological quality, we measured the diversity and abundance of macrobenthos assemblages. We detected p-p'DDE contamination, with concentrations higher than 3  $\mu\text{g kg}^{-1}$  in 16 out of 26 samples which were attributed to recent widespread use of DDT. The detection of sterols revealed faecal contamination. Significant sensitivity of the macrobenthos to contaminants was revealed, with possible implications on the overall mangrove vulnerability to climate change and on the provision of ecosystem services to local populations.

## Keywords

Mangrove; Contaminants; Macrobenthos; Cameroon; Wouri Estuary; Sediment.

48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

**Highlights**

- Contaminant and macrobenthonic survey was carried out in two Cameroonian mangrove forests around Douala city, Wouri estuary.
- Heavy metals, PCBs, PAHs, DDT and metabolites, and sterols were recorded in mangrove sediment.
- Crab and mollusc assemblages were assessed in both forests and were correlated with the presence of contaminants.
- Uncontrolled management of urban wastewater was among the most important factor of mangrove contamination.
- Integration of chemical and ecological data is required for a comprehensive risk assessment of peri-urban impacted mangrove forest.

## 1. Introduction

Mangroves in Africa cover over 3.2 million ha, corresponding to about 20% of their global coastline coverage, with approximately 1.5 million ha located along the Atlantic coast (Giri et al., 2011; Massó i Alemán et al., 2010; Spalding et al., 2010; UNEP, 2007). As a consequence of enormous anthropogenic pressure and multiple threats, Western African mangroves have declined by more than 25% over the past 25 years (Friess and Webb, 2014; Giri et al., 2011). Cameroon harbours approximately 2000 km<sup>2</sup> of mangroves, distributed along the coast of the Guinean gulf (Giri et al., 2011). Although mangroves contribute considerably to the social and economic well-being of the Cameroonian coastal inhabitants, their total surface area has decreased by 30% in 20 years (Spalding et al., 2010), mainly due to rapid and uncontrolled urbanization around Douala (Din et al., 2002; Ellison and Zouh, 2012; Nfotabong-Atheull et al., 2013). With a population of more than 2 million people, Douala is the largest city in Cameroon and exerts a huge pressure on the nearby mangroves, with uncontrolled sewage discharge detrimentally affecting the whole ecosystem (Simon and Raffaelli, 2012).

Douala is also one of the major shipping ports in the Guinea Gulf that serves the entire central Africa and refuels oil tankers to export locally extracted oil, another significant anthropogenic impact on the Wouri River estuary mangroves (Alemagi, 2007; Duke, 2016; Price et al., 2000; Walle, 1989). Due to the lack of policy regulation in the management of Cameroonian coastal ecosystems, sand mining and wood harvesting also play an important role in reducing mangrove biodiversity and provision of ecosystem services (Ellison and Zouh, 2012; Nfotabong-Atheull et al., 2011).

Although these multiple impacts threaten Wouri River estuary forests, the major socio-economic activity associated with mangroves for local people is in fact still artisanal fishing, with landings estimated between 76 and 106 tons per year (Gabche, 1997). Fisheries play a significant role in small-scale commercial activities and they are vital in providing a source of protein and income for coastal communities (Nfotabong-Atheull et al., 2009). Thus, the modification of both abundance and diversity of mangrove species and the deterioration of water quality, due to urban and industrial activities, will surely have detrimental consequences on the well-being of local communities (Alemagi, 2007; Nfotabong-Atheull et al., 2011, 2009). Last but not least, vulnerability to climate change, and especially to sea level rise, proved to be exacerbated by the high level of anthropogenic pressure on the Wouri River estuary mangroves (Ellison and Zouh, 2012). In particular, purported impacts of sea level rise on mangroves are (i) the increase of frequency and duration of the tidal inundation that may cause the death of the mangrove trees (exceeding the species-specific physiological thresholds; Ball, 1988), (ii) the impact on the inland fresh groundwater with saline intrusion and contaminants dispersal in the intertidal systems (Woodroffe et al., 2016), and (iii) the change of the topography and hydrology of the sediment (Lovelock et al., 2015).

Our aim was to perform the first baseline study on the ecological status and pollution of the strongly impacted Wouri River estuary mangroves, collecting data on both the presence of anthropogenic pollutants in sediments and the structure and diversity of macrobenthic populations as a proxy for healthy ecosystem functioning (Cannicci et al., 2009, 2008). To assess the level of chemical pollution, we targeted the major anthropogenic compounds usually found in peri-urban impacted mangrove forests world-wide, i.e. organochlorine compounds, such as DDT and its metabolites, phthalates such as bis(2-ethylhexyl)phthalate (DEHP), polycyclic aromatic hydrocarbons (PAHs), polychlorobiphenyls (PCBs), heavy metals and sterols (Bayen et al., 2016, 2005; Lewis et al., 2011; MacFarlane et al., 2007; Peters et al., 1997; Vane et al., 2009). Compounds such as coprostanol (5 $\beta$ (H)-Colestan-3  $\beta$  -ol) can be used in conjunction with other sterols to determine the relative abundance of sewage in sediments. Coprostanol, in particular, is a faecal sterol generated by microbial activity on cholesterol and is considered as a chemical marker of faecal contamination, especially from humans (Bull et al., 2002; Fattore et al., 1996; Mudge et al., 1999; Peng et al., 2005; Sherwin et al., 1993).

112 Diversity and abundance of crab and mollusc populations were recently shown to be key-  
113 determinants of the maintenance of mangrove ecosystem function and services (Cannicci et al.,  
114 2008; Duke et al., 2007; Lee, 2008), such as the provision of nursery sites for fish stocks which is of  
115 great importance for the local economy. Crabs and molluscs form an important link between  
116 primary detritus at the base of the food web and consumers at higher trophic levels (Sousa and  
117 Dangremond, 2011). By consuming litter, crabs can promote nutrient mineralization and recycling  
118 within the forest. Furthermore, their bioturbation activities undoubtedly alter the physico-chemical  
119 characteristics of soil (Kristensen, 2008) and enhance below-ground organic carbon retention  
120 (Andreetta et al., 2014). Finally, since mangrove macrobenthos diversity and functioning are known  
121 to be strongly impacted by contaminants (see Cannicci et al. 2009, Bartolini et al. 2011, Penha-  
122 Lopes et al. 2011 for east African mangrove benthos), their abundance and diversity is useful in  
123 assessing the degree of bioavailability of anthropogenic pollutants and the actual impact on the  
124 biological components.

## 125 126 **2. Material and Methods**

### 127 *2.1 Area description*

128 The study was carried out in two peri-urban mangrove forests located at different sites along the  
129 Wouri estuary: Wouri Bridge forest (4°4'19.10880''N; 9°42'5.81312'' E, hereafter WB) and Bois  
130 des Singes forest (4°0'49.67706''N; 9°40' 28.10325''E, hereafter BS), located north-west and  
131 south-east of Douala, respectively (fig. 1), with a distance between them of 11 km. Both of these  
132 stands are at about 10 hectares in extension and are largely affected by the uncontrolled expansion  
133 of urban areas due to the rapidly increasing population of Douala city (Simon and Raffaelli, 2012).  
134 Thus, they are representative sites to assess the possible presence of pollutants in peri-urban  
135 mangroves.

136 The climate of the region belongs to the Equatorial regime (Din and Baltzer, 2008), characterised  
137 by a long rainy season (March – November) and a short dry season (December – February). Heavy  
138 rainfall (approximately 4000 mm per year), stable high temperatures (annual average temperature is  
139 26.7°C) and high humidity throughout the year (approaching 100%) are typical for this region. The  
140 tidal regime is semi-diurnal with an average amplitude of 2.5 m. Soils are grey or black muds, of  
141 silty, sandy or clayey texture, derived from fluvial sediments relatively rich in organic matter with a  
142 high C:N ratio due to the reduced biological activity (Campo and Darius, 2004). Annual salinity  
143 variation in the region ranges between 0 and 20‰. During the long monsoon season, mangrove  
144 water salinity is consistently <10‰. During the dry season, salinity varies between 4 and 20‰ (Din  
145 and Baltzer, 2008).

146 According to a survey by Saenger & Bellan (1995), the floristic composition of Wouri Bridge  
147 forest, a 40 year old stand, is dominated by *Avicennia germinans*, *Rhizophora racemosa*, *R. mangle*  
148 and *R. harrisonii* and the mangrove associate *Pandanus* sp. Bois des Singes is an older stand and  
149 has a different floristic composition, represented only by the three *Rhizophora* species listed above.  
150 Hereafter, the belts studied in the Wouri Bridge forest will be referred to as the *Avicennia* belt,  
151 *Pandanus* belt and *Rhizophora* belt.

152 In these systems the faunal composition includes vertebrates, such as mammals like the sirenian  
153 species *Trichechus senegalensis* and several cetaceans. There are birds, of which particular  
154 significance is the high abundance of african skimmers, grey pranticoles, open-billed storks and  
155 common green shanks, reptiles such as four species of sea turtles (see Diop et al., 2014), and fishes  
156 (many of them of commercial importance), and a wide range of invertebrates, mainly crabs  
157 (belonging to the families Sesarmidae and Ocypodidae) and molluscs (extensively described by  
158 Ngo-Massou et al. 2012), which constitute the bulk of benthic diversity in the these mangrove  
159 ecosystem.

### 160 *2.2 Sediment Sampling*

162 A total of 20 sediment samples were collected in September 2009 in the two mangrove systems for  
163 trace metals and organic compounds analysis, choosing five random replicates in each forest. Five  
164 samples from BS and five from WB were taken from the upper layer of superficial sediment (0 – 10  
165 cm) and other five samples from BS and five from WB were taken from the layer underneath (11 –  
166 20 cm). The sediment cores were collected using an Eijkelpamp Multisampler™ piston corer (10  
167 cm diameter). The samples were then placed in glass jars, covered with aluminium foil and  
168 immediately transferred to a portable freezer and stored at -20° C until analysis.

## 169 170 2.3 Analytical methods

### 171 2.3.1 Solvents, chemicals and standards

172 The solvents used were acetone, hexane, dichloromethane and isooctane, obtained from Sigma  
173 Aldrich and Fluka Co., Steinheim, Germany. Standard reference materials for trace metals analysis  
174 were supplied by the Community Bureau of Reference Sample (BCR): Certified Reference  
175 Materials CRM 277 and CRM 320 and 142 R. Analytical standards for a mixture of PCBs (IUPAC  
176 nr. 28, 52, 101, 118, 138, 153, 180), a mixture of PAHs (anthracene, benzo[a] anthracene, benzo  
177 [j]k]fluoranthene, benzo [a] pyrene, benzo [ghi] perylene, chrysene , fluoranthene, indeno[1,2,3-  
178 cd]pyrene, phenanthrene, pyrene), bis(2-ethylhexyl)phthalate (DEHP) and the internal standards  
179 Anthracene-d<sub>10</sub> and Perylene-d<sub>12</sub> were purchased from Dr. Ehrenstorfer GmbH, Augsburg,  
180 Germany. Analytical sterol standards, Coprostan-3-ol , 5 α-Cholestan-3β-ol , Cholesterol and 5 β-  
181 Cholestan-3α-ol, analytical Bis(trimethylsilyl)trifluoroacetamide (BSTFA) with 1%  
182 trimethylchlorosilane (TMCS), used for sterols derivatisation, and analytical standards for 1,1,1-  
183 trichloro-2,2-di(4-chlorophenyl)ethane (DDT) and 1,1-bis-(4-chlorophenyl)-2,2-dichloroethene (p-  
184 p'DDE) were purchased from Sigma Aldrich and Fluka Co, Steinheim, Germany.

### 185 186 2.3.2 Trace metals analysis

187 All analyses were performed one month after sampling. Trace element concentrations of Mn, Pb,  
188 Cr, Cu, Zn, V and Ni were determined by inductively coupled plasma optical emission spectrometry  
189 (ICP OES, Perkin Elmer Optima 2100 DV spectrometry, Massachusetts, USA). Concentrations of  
190 Co, As, Se, Mo, Cd, Sn, Sb and Tl were determined by inductively coupled plasma mass  
191 spectrometry (ICP MS, Agilent Technologies mod. 7700x with Octapole Reaction System ORS,  
192 Santa Clara, USA) following aqua regia digestion according to Bettinelli et al. (2000). Mercury was  
193 determined by an automatic solid Hg analyser AMA 254. Certified and experimental values  
194 exhibited consistent values, recoveries ranging between 93 to 106% with repeatability better than  
195 8% using CRM 277 'Estuarine Sediment' and CRM 320 'River Sediment'. Certified soils and  
196 sediments were supplied by The Community Bureau of Reference Sample (BCR – IRMM, Joint  
197 Research Center, Retieseweg, B-2440, Geel, Belgium): CRM 142 'Light Sandy Soil', CRM 277  
198 'Estuarine Sediment' and CRM 320 'River Sediment'. Samples were handled according to the  
199 supplier's specifications. The MDL value was also calculated by analysing the blanks prepared on  
200 different working days. This parameter was assumed to be three times the blanks' standard  
201 deviation (See table S1 in the supplementary material).

### 202 203 2.3.3 Organic compounds analysis

204 For the extraction of PAHs (anthracene, benzo[a] anthracene, benzo [j]k]fluoranthene, benzo [a]  
205 pyrene, benzo [ghi] perylene, chrysene, fluoranthene, indeno[1,2,3- cd]pyrene, phenanthrene,  
206 pyrene, PCBs (IUPAC nr. 28, 52, 101, 118, 138, 153, 180), DEHP, DDT and its metabolite,  
207 samples were treated according to Zaccone et al. (2009). After extraction with Soxhlet using a  
208 hexane (80%) and acetone (20%) mixture and concentration of the extracts using a Buchi B-811  
209 Rotavapor, the obtained solutions were divided into two equal parts. A 5 ml aliquot of the extract  
210 was evaporated under a gentle flow of nitrogen, recovered with 0.5 mL of hexane containing the  
211 internal standards anthracene d<sub>10</sub> (1.14 mg l<sup>-1</sup>) and perylene d<sub>12</sub> (1.05 mg l<sup>-1</sup>), centrifuged and  
212 analyzed with GC-MS to determine the presence of benzo[j]k]fluoranthene (m/z 252),

213 benzo[a]pyrene (m/z 252), benzo [ghi]perylene (m/z 276), and indeno[1,2,3- cd]pyrene (m/z 276),  
214 DEHP (m/z 149 ), DDT (m/z 235) and p-p'DDE (m/z 246). The remaining 5 mL of the extracts  
215 were cleaned-up through a Florisil column, evaporated under a gentle flow of nitrogen, dissolved in  
216 0.5 mL of hexane containing the internal standards anthracene d<sub>10</sub> (1.14 mg l<sup>-1</sup>) and perylene d<sub>12</sub>  
217 (1.05 mg l<sup>-1</sup>), and then analysed to determine the presence of anthracene (m/z 178), chrysene (m/z  
218 228), benzo [a]anthracene (m/z 228), phenanthrene (m/z 178), pyrene (m/z 202) and fluoranthene  
219 (m/z 202), and of PCBs (m/z 256 for nr.28, m/z 292 for nr. 52, m/z 326 for nr. 101 and nr. 118, m/z  
220 360 for nr.138 and nr. 153 and m/z 394 for nr.180).

221 GC-MS analysis was performed according to Zaccone et al. (2009). Total Ion Monitoring (TIM)  
222 and Selected Ion Monitoring mode (SIM) were used for identification and quantification of  
223 substances. Overall concentration of PAHs and PCBs in sediment is the sum of the 12 PAHs and 7  
224 PCBs, respectively, analysed individually (mean of five samples for each site) expressed on a dry  
225 weight basis.

226 The sterols were analysed following the method proposed by Froehner et al. (2009), which includes  
227 Soxhlet extraction, clean-up with a silica-aluminium column, derivatisation and detection and  
228 quantification by GC-MS. The LODs were 0.2 µg kg<sup>-1</sup> for coprostan-3-ol and 0.4 µg kg<sup>-1</sup> for 5 α-  
229 Cholestan-3β-ol, Cholesterol and 5 β-Cholestan-3α-ol. The recoveries of sterols/stanols were  
230 between 65 and 80%.

231 The sediment total organic carbon (TOC) and total nitrogen (TN) in pore water were determined  
232 using the standard methods recommended by SSSA (Sparks et al., 1996).

#### 233 234 235 236 237 238 239 240 241 242 243 244 245 246 247 248 249 250 251 252 253 254 255 256 257 258 259 260 261 262 263 264 265 266 267 268 269 270 271 272 273 274 275 276 277 278 279 280 281 282 283 284 285 286 287 288 289 290 291 292 293 294 295 296 297 298 299 300 301 302 303 304 305 306 307 308 309 310 311 312 313 314 315 316 317 318 319 320 321 322 323 324 325 326 327 328 329 330 331 332 333 334 335 336 337 338 339 340 341 342 343 344 345 346 347 348 349 350 351 352 353 354 355 356 357 358 359 360 361 362 363 364 365 366 367 368 369 370 371 372 373 374 375 376 377 378 379 380 381 382 383 384 385 386 387 388 389 390 391 392 393 394 395 396 397 398 399 400 401 402 403 404 405 406 407 408 409 410 411 412 413 414 415 416 417 418 419 420 421 422 423 424 425 426 427 428 429 430 431 432 433 434 435 436 437 438 439 440 441 442 443 444 445 446 447 448 449 450 451 452 453 454 455 456 457 458 459 460 461 462 463 464 465 466 467 468 469 470 471 472 473 474 475 476 477 478 479 480 481 482 483 484 485 486 487 488 489 490 491 492 493 494 495 496 497 498 499 5002.3.4 Quality control

Calibration curves, prepared by dilution of stock solution with hexane, for PAHs and PCBs and  
DEHP were obtained at concentrations between 0.01 – 0.2 mg L<sup>-1</sup> and 0.1 and 2 mg L<sup>-1</sup>,  
respectively, using anthracene d<sub>10</sub> (1.14 mg L<sup>-1</sup>) and perylene d<sub>12</sub> (1.05 mg L<sup>-1</sup>) as internal  
standards. The calibration curves for p-p'DDE and DDT were created at concentrations between  
0.01 and 1 mg L<sup>-1</sup>, using anthracene d<sub>10</sub> (1.14 mg L<sup>-1</sup>) and perylene d<sub>12</sub> (1.05 mg L<sup>-1</sup>) as internal  
standards. Calibration curves for sterols were obtained at concentrations between 0.05 – 5 mg L<sup>-1</sup> by  
the dilution of stock solution with isooctane. Recovery experiments ( in triplicate) for PAH and  
PCBs were performed on a Certificate Reference Material (CRM) IMEP-21 obtained from the  
European Commission–JRC–IRMM with obtained values between 84 and 130% for PAHs and  
between 70 and 115% for PCBs and standard deviation, in all the cases, less than 20%. The limit of  
detection (LOD) was 2 µg kg<sup>-1</sup> for PAHs and 0.5 µg kg<sup>-1</sup> for PCBs. Due to the fact that DEHP was  
present in the extraction solvents and other materials used for extraction, three blank extractions  
were undertaken during the DEHP extraction process for all the samples. The DEHP sample  
concentrations were then corrected based on the daily blank extraction values. A Recovery  
experiment (in triplicate) for DEHP was performed on a sewage sludge sample by sample  
contamination at a concentration of 1 mg kg<sup>-1</sup>, with obtained values of 90% and standard deviation  
of 5%. The LOD for DEHP was 5 µg kg<sup>-1</sup> whereas for p-p'DDE it was 0.5 µg kg<sup>-1</sup>. The recoveries  
of sterols/stanols were between 65 and 80% with a standard deviation less than 5%, whereas the  
LODs were 0.2 µg kg<sup>-1</sup> for coprostan-3-ol and 0.4 µg kg<sup>-1</sup> for 5 α-Cholestan-3β-ol, Cholesterol  
and 5 β-Cholestan-3α-ol.

#### 2.4 Macrobenthos survey

In both forests, the surveys were carried out during three consecutive spring tides in September –  
October 2009. In each forest, two random transects (100–500 m apart) were established in each  
vegetation belt following a nested design. Along each transect, three 2 × 2 m<sup>2</sup> quadrats were  
randomly sampled to assess the abundance and density of the brachyuran and molluscan  
populations. The surveys were replicated for each spring tide period, at the same time of the day for  
the same time of observation in agreement with the methods described in Skov et al. (2002). Based

264 on the complexity of the habitat and the diverse behaviour of the study species, different sampling  
265 techniques were used to assess the abundance of the various groups of macrofauna. Due to their  
266 high densities, molluscs were counted in a sub-quadrat of  $50 \times 50 \text{ cm}^2$  placed within the sampling  
267 quadrat. Small sesarmids were counted visually throughout the quadrats. Large sesarmids were  
268 assessed by counting the number of operational burrows within the quadrats, since previous studies  
269 in South Africa and Kenya have clearly shown that these refuges are occupied by single crabs (Berti  
270 et al., 2008; Emmerson, 2001; Fratini et al., 2000; Skov et al., 2002). In order to refine the  
271 evaluation of crab and mollusc numbers, and due to the accumulation of leaf litter obscuring crabs,  
272 after observation for 1 hour in every quadrat we removed fallen leaves and logs to count the hidden  
273 specimens. Furthermore we measured temperature, pH and conductivity of the sediment water for  
274 each plot in each location using an Acorn pH 6 meter probe (Oakton Instruments).

## 275 276 277 2.5 Statistical analyses

278 A non-metric multidimensional scaling ordination (nMDS) was performed on the basis of a Bray-  
279 Curtis dissimilarity matrix calculated on untransformed data to visualize patterns of macrobenthic  
280 composition across sites. Furthermore, a PERMANOVA (Anderson et al., 2008) was used to test  
281 the null hypothesis of no differences in macrobenthos assemblages and temperature, pH and  
282 conductivity across the factor Site (fixed, orthogonal, two levels: WB – Wouri Bridge and BS –  
283 Bois des Singes), Belt (fixed, orthogonal three levels: *Rhizophora*, *Pandanus* and *Avicennia*) and  
284 Transect (random, nested in Site, 2 levels). PERMANOVA was also used to test the null hypothesis  
285 that there were no differences in contaminants, with the factors Site (fixed, orthogonal, two levels:  
286 WB – Wouri Bridge and BS – Bois des Singes), Belt (fixed, orthogonal three levels: *Rhizophora*,  
287 *Pandanus* and *Avicennia*) and Sampling Depth (fixed, orthogonal two levels: 0-10 and 11-20 cm).  
288 In the statistical analysis of contaminants, all values were normalised as performed by Dafforn et al.  
289 (2012), and Spearman's correlation was performed in order to eliminate covariate variables. For  
290 contaminants, the Euclidean distance was used to calculate the dissimilarity matrix. DistLM  
291 Analysis was performed to test the significant relationship between the ecological and  
292 anthropogenic factors and the macrobenthos assemblages.

## 293 294 3. Results

### 295 3.1 Sediment analysis

296 Concentrations of ten PAHs (fig. 2, supplementary table S2) and six PCBs congeners (table 1) were  
297 lower than  $300 \mu\text{g kg}^{-1}$  and  $20 \mu\text{g kg}^{-1}$  of sediment dry weight respectively. No statistical  
298 differences were recorded among sites, belts and sampling depths. Similarly, metals (fig. 2,  
299 supplementary table S3) and DEHP (table 2) concentrations were consistently lower than  $1500 \mu\text{g}$   
300  $\text{kg}^{-1}$ . DDT was absent, whereas its metabolite p-p'DDE (table 2) was found at concentrations higher  
301 than  $3 \mu\text{g kg}^{-1}$  in 16 samples. Furthermore, no statistical differences among the samples or between  
302 sites, belts and depths were observed. Cholesterol and 5  $\alpha$ -cholestan-3 $\beta$ -ol concentrations were  
303 lower than  $1000 \mu\text{g kg}^{-1}$  (fig. 2, supplementary table S4), while concentrations of coprostan-3-ol  
304 and 5  $\beta$ -cholestan-3 $\alpha$ -ol were lower than  $4000 \mu\text{g kg}^{-1}$  (with the exception of two samples). No  
305 statistical differences were detected among sites, belts and sampling depths. Interestingly,  
306 Coprostanol and 5  $\beta$ -cholestan-3 $\alpha$ -ol were not found in the deepest layers of sediment of any of the  
307 belts examined at the Wouri Bridge site.

308 Ratios between different sterols are presented in table 3 and they provide information concerning  
309 the source of contamination (according with Froehner et al. 2009), revealing widespread high  
310 disturbance in both forests.

### 311 312 3.2 Macrobenthic assemblages

59  
60  
61  
62  
63  
64  
65



313 The two forests were similar in terms of temperature, sediment water pH and sediment water  
314 conductivity (PERMANOVA, n=25, F=0.42, df=1,25, p > 0.05; table 4), as well as total N and total  
315 OC (PERMANOVA, n=28, F=0.89, df=2,28, p > 0.05; table 4)  
316 In accordance with the species listed by Ngo-Massou et al. (2012) and the reference list edited by  
317 Ng & Davie (2008) and Manning & Holthuis (1981), we individuated seven species of sesarimid and  
318 two species of mollusc inhabiting the forest. Within Sesarimidae, with an average density of 1.5  
319 individual per square meter, we identified *Perisesarma kamermanni*, *Perisesarma huzardi*,  
320 *Metagrapsus curvatum* and *Sesarma angolense* as burrowers and *Armases elegans* as a climber. The  
321 non-burrowing *Perisesarma alberti* and the potentially phytothelmic *Sesarma buettikoferi* (Fusi et  
322 al. unpublished data, fig. 4B), on the other hand, were more dense in the *Avicennia* belt of Wouri  
323 Bridge forest (with an average of about 4 individuals per square meter) the former and in the  
324 *Pandanus* belt in the same forest (almost 6 individuals per square meter) for the latter. Within  
325 molluscs, we recorded the presence of the Thiarid *Pachymelania fusca* in the two forests with a  
326 density of at about 400 specimens per square meter and the Potamidid *Tympanotonus radula* (fig.  
327 4A) found only in Bois de Singes forest with a density of at about 40 individuals per square meter.  
328 A significant difference in macrobenthos assemblages between Wouri Bridge and Bois des Singes  
329 forest was recorded (F = 25.655, p = 0.01, fig. 3; fig. 4). Specifically, Bois des Singe was  
330 characterized by the absence of *S. buettikoferi* and *S. angolense* and a more evenly distributed  
331 species density. In Wouri Bridge forest, a dominant species for each belt was observed: *P. alberti*  
332 was dominant in the *Avicennia* belt, *S. buettikoferi* in the *Pandanus* belt, while *P. alberti* and *P.*  
333 *kamermanni* were the two most abundant species in the *Rhizophora* belt. There was a notable  
334 absence of the gastropod *T. radula* throughout all the Wouri Bridge transects.

### 3.3 Macrobenthos assemblages and contaminants

337 The DistLM analysis shows a significant relationship between macrobenthos assemblage and  
338 sterols, metals, PAHs and C/N data (table 5; fig. 5a) and explains more than 90% of the total  
339 variation. In particular, 5  $\beta$ -cholestan-3 $\alpha$ -ol 1, Selenium, Chromium and Zinc explain the highest  
340 percentage of variation (20, 15, 8 and 7% respectively, p < 0.01). The two species of mollusc, the  
341 crabs *P. huzardi* and *P. alberti* appear to be most affected by the variation of the significant  
342 environmental variables cited above (fig. 5b).

## 4. Discussion

345 Mangroves in Cameroon still cover many hectares of estuaries, especially along the Wouri River  
346 where a complex system of channels and fens hinder access, and thus direct exploitation, mainly  
347 represent by logging for household and coal, and land claim for building new settlements.  
348 Nevertheless, the rapid development of Douala together with important commercial and trade  
349 activities, due to the presence of the harbour, have contributed to the city being a source of  
350 contaminants, which are spreading into nearby mangrove forests. We revealed in this study the  
351 presence of contaminants such as PCBs, PAHs, DEHP and heavy metals.

### *Effect of Sterols, p-p-DDE and heavy metals contamination on macrobenthic fauna*

354 In particular, p-p-DDE (DDT metabolite) was found in high concentrations close to the threshold  
355 admissible limit (Yang et al., 2007), as previously reported in other mangrove systems by Bodin et  
356 al. (2011) and Bhupander and Debapriya (2012). It is likely that this high level is related to recent  
357 intensive, and mainly uncontrolled, anti-malaria treatment in the area (Antonio-Nkondjio et al.,  
358 2011; Denison, 2013; Etang et al., 2007; Fossog Tene et al., 2013). Unfortunately, these compounds  
359 are reported to have a toxic effect on marine organisms (e.g. Bayarri et al. 2001, Mearns et al. 2014)  
360 and we strongly suggest that their high levels found in Wouri Bridge might be the reason for the  
361 total lack of *Tympanotonus radula* (fig 4, table 2) since the documented endocrine disrupting  
362 mechanism of p-p-DDE in molluscs (Matthiessen, 2008). Extremely rapid urbanisation has resulted  
363 in a growing urban population that has colonised areas within well-established rainforest and



364 mangrove forests. In these areas, settlements consist of rudimentary housing with uncontrolled  
365 discharge of untreated sewage and wastewater into the forests (Nfotabong-Atheull et al., 2011;  
366 Simon and Raffaelli, 2012). This is likely the cause of the presence of sterols detected with a high  
367 ratio of (Coprostanol + epicoprostanol)/  $\Sigma$  Total Stanols that indicates a serious level of sediment  
368 sewage contamination (Froehner et al., 2009; Gern and Lana, 2013). Specifically, we identified high  
369 contamination in 10 of the 26 samples (4 in Wouri Bridge and 6 in Bois des Singes). This  
370 contamination is determined not only by the uncontrolled urbanization taking place in Douala, but  
371 also by the lack of any wastewater treatment management in the city. Indeed, the area nearby Bois  
372 de Singes is highly affected by municipality wastewater discharge, with trucks (personal  
373 observation) releasing tons of untreated wastewater directly into the mangrove, also witnessed  
374 during the survey. This activity is also a good explanation of the fact that we found more highly  
375 contaminated samples in Bois de Singes than Wouri Bridge. Through the macrobenthic survey, we  
376 were able to record highly biodiverse and structured macrobenthic communities in both forests.  
377 However, we recorded two significantly different patterns of macrobenthic assemblage at the two  
378 sites, mainly due to the absence of *T. radula* (Potamididae) and *P. huzardi* (Sesarmidae) in Wouri  
379 Bridge forest, while *S. buettikoferi* was not found in Bois de Singes forest (Fig. 4). Our statistical  
380 analyses found 5  $\beta$ -cholestan-3 $\alpha$ -ol (Gern and Lana, 2013) and four heavy metals, As (Beltman et  
381 al., 1999), Se (Hamilton, 2004), Cr (Lewis et al., 2001) and Zn (Ellis et al., 2004; Schaffelke et al.,  
382 2005), to be the main drivers of differences in crab assemblage (See Fig. 5 and table 5). Indeed,  
383 they are known to be among the more important compounds responsible to disrupt the physiology  
384 of marine species (e.g. Mello and Nayak, 2016 and reference above). If we consider that the main  
385 environmental features, including the tidal regime, were similar between sites and belts, we  
386 hypothesize that these contaminant levels could play a major role shaping the macrobenthos  
387 distribution and density as a result of their differential sensitivity to pollutants and their  
388 concentration. Such a selective ecological effect has been largely described for molluscs in east  
389 African mangrove systems exposed to sewage (Cannicci et al., 2009; Penha-Lopes et al., 2010).

390  
391 *Implications of anthropogenic contamination on mangrove ecosystem*

392 Currently, the interpretation of biological responses as a consequence of contamination remains  
393 complex. A major reason being that organisms in the field are exposed to multiple stressors under  
394 dynamic conditions (e.g. variable micro- and macronutrient loads, changing climatic conditions,  
395 multiple contaminants, tidal cycles and salinity), and potential additive, synergistic or antagonistic  
396 responses to these stressors may occur (Bayen, 2012). Indeed, changes in the diversity/structure of  
397 mangrove ecosystems have been reported as a response to chemical pollution (e.g. Mohamed et al.  
398 2008), which has also been linked to a decline in some populations such as mangrove oysters and  
399 snails (e.g. Roach & Wilson 2009) and molluscs (Cannicci et al., 2009). Kulkarni et al. (2010)  
400 reported low biodiversity indices associated with a low water quality index in mangrove ecosystems  
401 in India. The overall impact of pollution, however, appears to be complex. For example, the patterns  
402 of diversity and species composition recorded in various mangrove forests highly impacted by  
403 humans in Indonesia did not clearly correlate with the impact investigated (Geist et al., 2012).  
404 Moreover, discharge of domestic sewage at low levels caused an increase in crab population size in  
405 east Africa (Cannicci et al., 2009; Penha-Lopes et al., 2011) and did not affect the macrobenthic  
406 communities in Hong Kong (Wong et al., 1997; Yu et al., 1997). Our results indicate that multiple  
407 anthropogenic stressors, and in particular heavy loads of wastewater, although not resulting in the  
408 hypothesised depletion of crab abundance in Wouri River estuary mangroves, can shape their  
409 community composition. The data suggest that contaminant loadings in Wouri river mangroves,  
410 affecting the distribution of macrobenthonic species, could lead to the type of cryptic ecological  
411 degradation (*sensu* Dahdouh-Guebas et al. 2005) shown by Bartolini et al. (2011), who documented  
412 an inverse relationship between the increased biomass of fiddler crabs and their overall engineering  
413 function, thus affecting the whole mangrove ecosystem.

414 The sterols level recorded in this study strongly indicates a more large contamination by sewage  
415 and, in particular, by a first stages of human faecal contamination, as recorded in other studies (i.e.  
416 Fernandes et al. 1999, Silva & Madureira 2012). This can have an important effect on the ecology  
417 of macrobenthic species (Frena et al., 2016) by changing the chemistry of the sediment where they  
418 live dramatically enhancing bacterial activity (Dheenan et al., 2016). The subtle faecal  
419 contamination could triggering possible ecological bottom-up effect with relevant ecological  
420 consequences on the overall system (Hamilton, 2004), through the modification of secondary  
421 consumers primary feeding sources, such as microbenthic (Isobe et al., 2004) or infaunal  
422 macrobenthic communities (Moon et al., 2008). Together with the presence of PCBs, PAHs and  
423 heavy metals, chromium in particular, raises at least two important concerns. The first is that  
424 mangrove sediment receiving wastewater does accumulate a remarkably higher level of long-term  
425 contaminants (Tam and Wong, 1995). The second is that they could potentially spread through the  
426 whole mangrove area (Agoramoorthy et al., 2008; Yi et al., 2011; Zhou et al., 2007), ultimately  
427 affecting, through the trophic chain, the secondary and tertiary consumers which are consumed by  
428 the local population. Therefore, they potentially represent a health risk.

## 429 430 **5. Conclusions**

431 To our knowledge, this study is the first to investigate the contamination of the Cameroon estuarine  
432 and marine environment with Persistent Organic Pollutants (POPs). It contributes to the scarce data  
433 and literature on this subject in African countries, which has mainly focused on public health and  
434 resistance implications from POPs use. Our data clearly show that the main source of contamination  
435 in the mangrove forests surrounding Douala is represented by uncontrolled discharge of urban  
436 wastewater and the persistent, illegal and indiscriminate use of DDT. These contaminants, together  
437 with four specific heavy metals (As, Cr, Zn, Se) seem to affect the macrobenthonic assemblage of  
438 the two study sites, suggesting that Douala peri-urban mangrove is subjected to a complex  
439 patchwork of contamination. This documented inflow pollution has serious implications for  
440 ecosystem functioning and public health. Therefore we emphasize the necessity to prioritise water  
441 quality monitoring and the development of public policies for the wastewater management.  
442 Additionally, as highlighted in many studies, human pollution is likely to impair the provision of  
443 critical mangrove ecosystem services which are relied upon by local communities. Integrated  
444 assessment of macrobenthic assemblages should be considered as a method to detect early  
445 contamination patterns, as suggested by our results and confirmed by several other studies. Hence,  
446 the present data provide a baseline for further development and environmental management  
447 oriented towards anthropogenic pollution by POPs in West Africa, also in the view of monitoring  
448 and reducing human impact to mitigate vulnerability of mangroves to the fast climate change.

449  
450

451  
452  
453  
454  
455  
456  
457  
458  
459  
460  
461  
462  
463  
464  
465

450  
451  
452  
453  
454  
455  
456  
457  
458  
459  
460  
461  
462  
463  
464  
465  
466  
467  
468  
469  
470  
471  
472  
473  
474  
475  
476  
477  
478  
479  
480  
481  
482  
483  
484  
485  
486  
487  
488  
489  
490  
491  
492  
493  
494  
495  
496  
497  
498  
499  
500  
501  
502  
503  
504  
505  
506  
507  
508  
509  
510  
511  
512  
513  
514  
515  
516  
517  
518  
519  
520  
521  
522  
523  
524  
525  
526  
527  
528  
529  
530  
531  
532  
533  
534  
535  
536  
537  
538  
539  
540  
541  
542  
543  
544  
545  
546  
547  
548  
549  
550  
551  
552  
553  
554  
555  
556  
557  
558  
559  
560  
561  
562  
563  
564  
565

## Acknowledgements

The study was supported by the Cameroon Oil Transportation Company (COTCO), the SP3-People (Marie Curie) IRSES Project CREC (no. 247514), the SC research funds from MIUR (ex 60%) and supported by King Abdullah University of Science and Technology (baseline research funds to DD). We thank Adolphe Nfotabong-Atheull, Vanessa Ngo-Massou and Joseph Bayi for invaluable help during field sampling. We are grateful to Ilaria Marchi for design assistance and Pierluisa Fantini for invaluable laboratory work. Thanks to Jenny Marie Booth for English revision of this manuscript.

## References

- Agoramoorthy, G., Chen, F., Hsu, M.J., 2008. Threat of heavy metal pollution in halophytic and mangrove plants of Tamil Nadu, India. *Environ. Pollut.* 155, 320–326. doi:10.1016/j.envpol.2007.11.011
- Alemagi, D., 2007. The oil industry along the Atlantic coast of Cameroon: Assessing impacts and possible solutions. *Resour. Policy* 32, 135–145. doi:10.1016/j.resourpol.2007.08.007
- Anderson, M.J., Godley, R.N., Clarke, K.R., 2008. PERMANOVA+ for PRIMER: guide to software and statistical methods. PRIMER-E Ltd., Plymouth.
- Andreetta, A., Fusi, M., Cameldi, I., Cimò, F., Carnicelli, S., Cannicci, S., 2014. Mangrove carbon sink. Do burrowing crabs contribute to sediment carbon storage? Evidence from a Kenyan mangrove system. *J. Sea Res.* 85, 524–533. doi:10.1016/j.seares.2013.08.010
- Antonio-Nkondjio, C., Fossog, B.T., Ndo, C., Djantio, B.M., Togouet, S.Z., Awono-Ambene, P., Costantini, C., Wondji, C.S., Ranson, H., 2011. Anopheles gambiae distribution and insecticide resistance in the cities of Douala and Yaoundé (Cameroon): influence of urban agriculture and pollution. *Malar. J.* 10, 154. doi:10.1186/1475-2875-10-154
- Ball, M.C., 1988. Ecophysiology of mangroves. *Trees* 2, 129–142. doi:10.1007/BF00196018
- Bartolini, F., Cimò, F., Fusi, M., Dahdouh-Guebas, F., Lopes, G.P., Cannicci, S., 2011. The effect of sewage discharge on the ecosystem engineering activities of two East African fiddler crab species: consequences for mangrove ecosystem functioning. *Mar. Environ. Res.* 71, 53–61. doi:10.1016/j.marenvres.2010.10.002
- Bayarri, S., Baldassarri, L.T., Iacovella, N., Ferrara, F., Domenico, A.D., 2001. PCDDs, PCDFs, PCBs and DDE in edible marine species from the Adriatic Sea. *Chemosphere* 43, 601–610. doi:10.1016/S0045-6535(00)00412-4
- Bayen, S., 2012. Occurrence, bioavailability and toxic effects of trace metals and organic contaminants in mangrove ecosystems: A review. *Environ. Int.* 48, 84–101. doi:10.1016/j.envint.2012.07.008
- Bayen, S., Estrada, E., Juhel, G., Lee, W., Kelly, B., 2016. Pharmaceutically active compounds and endocrine disrupting chemicals in water, sediments and mollusks in mangrove ecosystems from Singapore. *Mar. Pollut. Bull.*
- Bayen, S., Wurl, O., Karuppiah, S., Sivasothi, N., Lee, H.K., Obbard, J.P., 2005. Persistent organic pollutants in mangrove food webs in Singapore. *Chemosphere* 61, 303–313. doi:10.1016/j.chemosphere.2005.02.097
- Beltman, D.J., Clements, W.H., Lipton, J., Cacula, D., 1999. Benthic invertebrate metals exposure, accumulation, and community-level effects downstream from a hard-rock mine site. *Environ. Toxicol. Chem.* 18, 299–307. doi:10.1897/1551-5028(1999)018<0299:bimeaa>2.3.co;2

- 495 Berti, R., Cannicci, S., Fabbroni, S., Innocenti, G., 2008. Notes on the structure and the use of  
496 Neosarmatium meinerti and Cardisoma carnifex burrows in a Kenyan mangrove swamp  
497 (Decapoda Brachyura). *Ethol. Ecol. Evol.* 20, 101–113.  
3
- 498 Bettinelli, M., Beone, G., Spezia, S., Baffi, C., 2000. Determination of heavy metals in soils and  
499 sediments by microwave-assisted digestion and inductively coupled plasma optical emission  
500 spectrometry analysis. *Anal. Chim. Acta* 424, 289–296. doi:10.1016/S0003-2670(00)01123-5
- 501 Bhupander, K., Debapriya, M., 2012. Eco-Toxicological Risk Assessment of HCH , DDT and their  
502 Possible Sources by Isomeric Ratio Distribution in Sediments from Sundarban Mangrove  
503 Ecosystem in Bay of Bengal , India. *J. Environ. Earth Sci.* 2, 58–71.
- 504 Bodin, N., N’Gom Ka, R., Le Loc’h, F., Raffray, J., Budzinski, H., Peluhet, L., Tito de Moraes, L.,  
505 2011. Are exploited mangrove molluscs exposed to Persistent Organic Pollutant contamination  
506 in Senegal, West Africa? *Chemosphere* 84, 318–327. doi:10.1016/j.chemosphere.2011.04.012
- 507 Bull, I.D., Lockheart, M.J., Elhmmali, M.M., Roberts, D.J., Evershed, R.P., 2002. The origin of  
508 faeces by means of biomarker detection. *Environ. Int.* 27, 647–654. doi:10.1016/S0160-  
509 4120(01)00124-6  
20
- 510 Campo, E. Van, Darius, M., 2004. Mangrove palynology in recent marine sediments off Cameroon.  
511 *Mar. Geol.* 208, 315–330. doi:10.1016/j.margeo.2004.04.014  
22  
23
- 512 Cannicci, S., Bartolini, F., Dahdouh-Guebas, F., Fratini, S., Litulo, C., Macia, A., Mrabu, E.J.,  
513 Penha-Lopes, G., Paula, J., 2009. Effects of urban wastewater on crab and mollusc  
514 assemblages in equatorial and subtropical mangroves of East Africa. *Estuar. Coast. Shelf Sci.*  
515 84, 305–317. doi:10.1016/j.ecss.2009.04.021  
25  
26  
27
- 516 Cannicci, S., Burrows, D., Fratini, S., Smith, T.J., Offenber, J., Dahdouh-Guebas, F., 2008. Faunal  
517 impact on vegetation structure and ecosystem function in mangrove forests: A review. *Aquat.*  
518 *Bot.* 89, 186–200. doi:10.1016/j.aquabot.2008.01.009  
31  
33
- 519 Dafforn, K. a., Simpson, S.L., Kelaher, B.P., Clark, G.F., Komyakova, V., Wong, C.K.C., Johnston,  
520 E.L., 2012. The challenge of choosing environmental indicators of anthropogenic impacts in  
521 estuaries. *Environ. Pollut.* 163, 207–217. doi:10.1016/j.envpol.2011.12.029  
35  
37
- 522 Dahdouh-Guebas, F., Hettiarachchi, S., Lo Seen, D., Batelaan, O., Sooriyachchi, S., Jayatissa,  
523 L.P., Koedam, N., 2005. Transitions in Ancient Inland Freshwater Resource Management in  
524 Sri Lanka Affect Biota and Human Populations in and around Coastal Lagoons. *Curr. Biol.* 15,  
525 579–586. doi:10.1016/j.cub.2005.01.053  
38  
39  
40  
41
- 526 Denison, L., 2013. Stockholm Convention on Persistent Organic Pollutants. *Air Qual. Clim. Chang.*  
527 2.  
43  
44
- 528 Dheenan, P.S., Jha, D.K., Das, A.K., Vinithkumar, N.V., Devi, M.P., Kirubakaran, R., 2016.  
529 Geographic information systems and multivariate analysis to evaluate fecal bacterial pollution  
530 in coastal waters of Andaman, India. *Environ. Pollut.* 214, 45–53.  
531 doi:10.1016/j.envpol.2016.03.065  
46  
50  
51
- 532 Din, N., Baltzer, F., 2008. Richesse Floristique et Evolution des mangroves de l’Estuaire du  
533 Cameroun. *African Geosci. Rev.* 2, 119–130.  
53  
54
- 534 Din, N., Priso, R.J., Kenne, M., Ngollo, D.E., Blasco, F., 2002. Early growth stages and natural  
535 regeneration of *Avicennia germinans* (L.) Stearn in the Wouri estuarine mangroves (Douala-  
536 Cameroon). *Wetl. Ecol. Manag.* 10, 461–472. doi:10.1023/A:1021351707822  
53  
56  
57  
58
- 59 Diop, S., Barousseau, J., Descamps, C., 2014. The Land/Ocean Interactions in the Coastal Zone of  
60 West and Central Africa, Springer, *Estuaries of the World*. Springer International Publishing,  
61  
62  
63  
64  
65

539 Cham. doi:10.1007/978-3-319-06388-1

540 Duke, N., Meynecke, J.O., Dittmann, S., Ellison, A.M., Anger, K., Berger, U., Cannicci, S., Diele,  
541 K., Ewel, K.C., Field, C.D., Koedam, N., Lee, S.Y., Marchand, C., Nordhaus, I., Dahdouh-  
542 Guebas, F., 2007. A World Without Mangroves ? Science (80-. ). 317, 41–43.  
543 doi:10.1126/science.317.5834.41b

544 Duke, N.C., 2016. Oil spill impacts on mangroves: a global review with recommendations. Mar.  
545 Pollut. Bull.

546 Ellis, J., Nicholls, P., Craggs, R., Hofstra, D., Hewitt, J., 2004. Effects of terrigenous sedimentation  
547 on mangrove physiology and associated macrobenthic communities. Mar. Ecol. Prog. Ser. 270,  
548 71–82. doi:10.3354/meps270071

549 Ellison, J.C., Zouh, I., 2012. Vulnerability to Climate Change of Mangroves: Assessment from  
550 Cameroon, Central Africa. Biology (Basel). 1, 617–638. doi:10.3390/biology1030617

551 Emmerson, W.D., 2001. Aspects of the population dynamics of *Neosarmatium meinerti* at Mgazana  
552 , a warm temperate mangrove swamp in the East Cape , South Africa , investigated using an  
553 indirect method 221–229.

554 Etang, J., Manga, L., Toto, J., Guillet, P., Fondjo, E., Etang, J., Manga, L., Toto, J., Guillet, P.,  
555 Fondjo, E., 2007. Spectrum of metabolic-based resistance to DDT and pyrethroids in  
556 *Anopheles gambiae* s . l . populations from Cameroon Spectrum of metabolic-based resistance  
557 to DDT and pyrethroids in *Anopheles gambiae* s . l . populations from Cameroon. J. Vector  
558 Ecol. 32, 123–133.

559 Fattore, E., Benfenati, E., Marelli, R., Cools, E., Fanelli, R., 1996. Sterols in sediment samples from  
560 Venice Lagoon, Italy. Chemosphere 33, 2383–2393. doi:10.1016/S0045-6535(96)00340-2

561 Fernandes, M.B., Sicre, M. a., Cardoso, J.N., Macêdo, S.J., 1999. Sedimentary 4-desmethyl sterols  
562 and n-alkanols in an eutrophic urban estuary, Capibaribe River, Brazil. Sci. Total Environ.  
563 231, 1–16. doi:10.1016/S0048-9697(99)00077-7

564 Fossog Tene, B., Poupardin, R., Costantini, C., Awono-Ambene, P., Wondji, C.S., Ranson, H.,  
565 Antonio-Nkondjio, C., 2013. Resistance to DDT in an Urban Setting: Common Mechanisms  
566 Implicated in Both M and S Forms of *Anopheles gambiae* in the City of Yaoundé?? Cameroon.  
567 PLoS One 8. doi:10.1371/journal.pone.0061408

568 Fratini, S., Cannicci, S., Vannini, M., 2000. Competition and interaction between *Neosarmatium*  
569 *smithi* (Crustacea : Grapsidae) and *Terebralia palustris* (Mollusca : Gastropoda) in a Kenyan  
570 mangrove. Mar. Biol. 137, 309–316.

571 Frena, M., Bataglion, G.A., Tonietto, A.E., Eberlin, M.N., Alexandre, M.R., Madureira, L.A.S.,  
572 2016. Assessment of anthropogenic contamination with sterol markers in surface sediments of  
573 a tropical estuary (Itajaí-Açu, Brazil). Sci. Total Environ. 544, 432–438.  
574 doi:10.1016/j.scitotenv.2015.11.137

575 Friess, D.A., Webb, E.L., 2014. Variability in mangrove change estimates and implications for the  
576 assessment of ecosystem service provision. Glob. Ecol. Biogeogr. 23, 715–725.  
577 doi:10.1111/geb.12140

578 Froehner, S., Martins, R.F., Errera, M.R., 2009. Assessment of fecal sterols in Barigui River  
579 sediments in Curitiba, Brazil. Environ. Monit. Assess. 157, 591–600. doi:10.1007/s10661-008-  
580 0559-0

581 Gabche, C.E., 1997. An appraisal of fisheries activities and evaluation of economic potential of the  
582 fish trade in the Douala–Edea reserve–Cameroon, Cameroon W. ed. Yaoundé, Cameroon.

- 583 Geist, S.J., Nordhaus, I., Hinrichs, S., 2012. Occurrence of species-rich crab fauna in a human-  
584 impacted mangrove forest questions the application of community analysis as an  
585 environmental assessment tool. *Estuar. Coast. Shelf Sci.* 96, 69–80.  
586 doi:10.1016/j.ecss.2011.10.002
- 587 Gern, F.R., Lana, P. da C., 2013. Reciprocal experimental transplantations to assess effects of  
588 organic enrichment on the recolonization of benthic macrofauna in a subtropical estuary. *Mar.*  
589 *Pollut. Bull.* 67, 107–120. doi:10.1016/j.marpolbul.2012.11.026
- 590 Giri, C., Ochieng, E., Tieszen, L.L., Zhu, Z., Singh, a., Loveland, T., Masek, J., Duke, N., 2011.  
591 Status and distribution of mangrove forests of the world using earth observation satellite data.  
592 *Glob. Ecol. Biogeogr.* 20, 154–159. doi:10.1111/j.1466-8238.2010.00584.x
- 593 Hamilton, S.J., 2004. Review of selenium toxicity in the aquatic food chain. *Sci. Total Environ.*  
594 326, 1–31. doi:10.1016/j.scitotenv.2004.01.019
- 595 Isobe, K.O., Tarao, M., Chiem, N.H., Minh, L.Y., 2004. Effect of Environmental Factors on the  
596 Relationship between Concentrations of Coprostanol and Fecal Indicator Bacteria in Tropical (   
597 Mekong Delta ) and Temperate ( Tokyo ) Freshwaters. *Appl. Environ. Microbiol.* 70, 814–821.  
598 doi:10.1128/AEM.70.2.814
- 599 Kristensen, E., 2008. Mangrove crabs as ecosystem engineers; with emphasis on sediment  
600 processes. *J. Sea Res.* 59, 30–43. doi:10.1016/j.seares.2007.05.004
- 601 Kulkarni, V.A., Jagtap, T.G., Mhalsekar, N.M., Naik, A.N., 2010. Biological and environmental  
602 characteristics of mangrove habitats from Manori creek, West Coast, India. *Environ. Monit.*  
603 *Assess.* 168, 587–596. doi:10.1007/s10661-009-1136-x
- 604 Lee, S.Y., 2008. Mangrove macrobenthos: Assemblages, services, and linkages. *J. Sea Res.* 59, 16–  
605 29. doi:10.1016/j.seares.2007.05.002
- 606 Lewis, M., Pryor, R., Wilking, L., 2011. Fate and effects of anthropogenic chemicals in mangrove  
607 ecosystems: A review. *Environ. Pollut.* 159, 2328–2346. doi:10.1016/j.envpol.2011.04.027
- 608 Lewis, M.A., Weber, D.E., Stanley, R.S., Moore, J.C., 2001. Dredging impact on an urbanized  
609 Florida bayou: Effects on benthos and algal-periphyton. *Environ. Pollut.* 115, 161–171.  
610 doi:10.1016/S0269-7491(01)00118-X
- 611 Lovelock, C.E., Cahoon, D.R., Friess, D.A., Guntenspergen, G.R., Krauss, K.W., Reef, R., Rogers,  
612 K., Saunders, M.L., Sidik, F., Swales, A., Saintilan, N., Thuyen, L.X., Triet, T., 2015. The  
613 vulnerability of Indo-Pacific mangrove forests to sea-level rise. *Nature* 526, 559–U217.  
614 doi:10.1038/nature15538
- 615 MacFarlane, G.R., Koller, C.E., Blomberg, S.P., 2007. Accumulation and partitioning of heavy  
616 metals in mangroves: A synthesis of field-based studies. *Chemosphere* 69, 1454–1464.  
617 doi:10.1016/j.chemosphere.2007.04.059
- 618 Manning, R.B., Holthuis, L.B., 1981. West African Brachyuran Crabs (Crustacea: Decapoda).  
619 *Smithson. Contrib. to Zool.* 1–396. doi:10.5479/si.00810282.306
- 620 Massó i Alemán, S., Bourgeois, C., Appeltans, W., Vanhoorne, B., De Hauwere, N., Stoffelen, P.,  
621 Heughebaert, A., Dahdouh-Guebas, F., 2010. The “Mangrove Reference Database and  
622 Herbarium.” *Plant Ecol. Evol.* 143, 225–232. doi:10.5091/plecevo.2010.439
- 623 Matthiessen, P., 2008. An assessment of endocrine disruption in mollusks and the potential for  
624 developing internationally standardized mollusk life cycle test guidelines. *Integr. Environ.*  
625 *Assess. Manag.* 4, 274–284. doi:10.1897/IEAM\_2008-003.1

- 626 Mearns, A.J., Reish, D.J., Oshida, P.S., Ginn, T., Rempel-Hester, M.A., Arthur, C., Rutherford, N.,  
627 2014. Effects of Pollution on Marine Organisms. *Water Environ. Res.* 86, 1869–1954.  
628 doi:10.2175/106143012X13407275695751  
3
- 629 Mello, C.A.N., Nayak, G.N., 2016. Assessment of metal enrichment and their bioavailability in  
630 sediment and bioaccumulation by mangrove plant pneumatophores in a tropical ( Zuari )  
631 estuary , west coast of India. *Mar. P In press.* doi:10.1016/j.marpolbul.2016.06.0590025-326X
- 632 Mohamed, M.O.S., Neukermans, G., Kairo, J.G., Dahdouh-Guebas, F., Koedam, N., 2008.  
633 Mangrove forests in a peri-urban setting: the case of Mombasa (Kenya). *Wetl. Ecol. Manag.*  
634 17, 243–255. doi:10.1007/s11273-008-9104-8
- 635 Moon, H.B., Yoon, S.P., Jung, R.H., Choi, M., 2008. Wastewater treatment plants (WWTPs) as a  
636 source of sediment contamination by toxic organic pollutants and fecal sterols in a semi-  
637 enclosed bay in Korea. *Chemosphere* 73, 880–889. doi:10.1016/j.chemosphere.2008.07.038  
16
- 638 Mudge, S.M., Bebianno, M.J. a F., East, J. a., Barreira, L. a., 1999. Sterols in the Ria Formosa  
639 lagoon, Portugal. *Water Res.* 33, 1038–1048. doi:10.1016/S0043-1354(98)00283-8  
19
- 640 Nfotabong-Atheull, A., Din, N., Dahdouh-Guebas, F., 2013. Qualitative and Quantitative  
641 Characterization of Mangrove Vegetation Structure and Dynamics in a Peri-urban Setting of  
642 Douala (Cameroon): An Approach Using Air-Borne Imagery. *Estuaries and Coasts* 36, 1181–  
643 1192. doi:10.1007/s12237-013-9638-8
- 644 Nfotabong-Atheull, A., Din, N., Essomè Koum, L.G., Satyanarayana, B., Koedam, N., Dahdouh-  
645 Guebas, F., 2011. Assessing forest products usage and local residents' perception of  
646 environmental changes in peri-urban and rural mangroves of Cameroon, Central Africa. *J.*  
647 *Ethnobiol. Ethnomed.* 7, 41. doi:10.1186/1746-4269-7-41  
30
- 648 Nfotabong-Atheull, A., Din, N., Longonje, S.N., Koedam, N., Dahdouh-Guebas, F., 2009.  
649 Commercial activities and subsistence utilization of mangrove forests around the Wouri  
650 estuary and the Douala-Edea reserve (Cameroon). *J. Ethnobiol. Ethnomed.* 5, 35.  
651 doi:10.1186/1746-4269-5-35
- 652 Ng, P.K.L., Davie, P.J.F., Guinot, D., 2008. *Systema Brachyurorum : Part I . an Annotated*  
653 *Checklist of Extant Brachyuran Crabs of the World.* *Raffles Bull. Zool.* 17, 1–286.
- 654 Ngo-Massou, V., Essome-Koum, G., Ngollo-Dina, E., Din, N., 2012. Composition of macrobenthos  
655 in the Wouri River estuary mangrove, Douala, Cameroon. *African J. Mar. Sci.* 34, 349–360.  
656 doi:10.2989/1814232X.2012.725288  
43
- 657 Peng, X., Zhang, G., Mai, B., Hu, J., Li, K., Wang, Z., 2005. Tracing anthropogenic contamination  
658 in the Pearl River estuarine and marine environment of South China Sea using sterols and  
659 other organic molecular markers. *Mar. Pollut. Bull.* 50, 856–65.  
660 doi:10.1016/j.marpolbul.2005.02.031  
47
- 661 Penha-Lopes, G., Kristensen, E., Flindt, M., Mangion, P., Bouillon, S., Paula, J., 2010. The role of  
662 biogenic structures on the biogeochemical functioning of mangrove constructed wetlands  
663 sediments – A mesocosm approach. *Mar. Pollut. Bull.* 60, 560–572.  
664 doi:10.1016/j.marpolbul.2009.11.008  
54
- 665 Penha-Lopes, G., Torres, P., Cannicci, S., Narciso, L., Paula, J., 2011. Monitoring anthropogenic  
666 sewage pollution on mangrove creeks in southern Mozambique: a test of *Palaemon concinnus*  
667 Dana, 1852 (Palaemonidae) as a biological indicator. *Environ. Pollut.* 159, 636–45.  
668 doi:10.1016/j.envpol.2010.09.029  
58
- 669 Peters, E.C., Gassman, N.J., Firman, J.C., Richmond, R.H., Power, E. a, 1997. *Ecotoxicology of*  
61



- 670 tropical marine ecosystems. *Environ. Toxicol. Chem.* 16, 12–40. doi:10.1002/etc.5620160103
- 671 Price, A., Klaus, A., Sheppard, C.R.C., Abbiss, M.A., Kofani, M., Webster, G., 2000.  
672 Environmental and bioeconomic characterisation of coastal and marine systems of Cameroon,  
673 including risk implications of the Chad–Cameroon pipeline project. *Aquat. Ecosyst. Heal.*  
674 *Manag.* 3, 137–161. doi:10.1016/S1463-4988(99)00072-X
- 675 Roach, A.C., Wilson, S.P., 2009. Ecological impacts of tributyltin on estuarine communities in the  
676 Hastings River, NSW Australia. *Mar. Pollut. Bull.* 58, 1780–1786.  
677 doi:10.1016/j.marpolbul.2009.08.021
- 678 Saenger, P., Bellan, M.F., 1995. *The Mangrove Vegetation of the Atlantic Coast of Africa.*  
679 Universite’ de Toulouse Press, Toulouse.
- 680 Schaffelke, B., Mellors, J., Duke, N.C., 2005. Water quality in the Great Barrier Reef region:  
681 Responses of mangrove, seagrass and macroalgal communities. *Mar. Pollut. Bull.* 51, 279–  
682 296. doi:10.1016/j.marpolbul.2004.10.025
- 683 Sherwin, M.R., Van Vleet, E.S., Fossato, V.U., Dolci, F., 1993. Coprostanol (5 $\beta$ -cholestan-3 $\beta$ -ol) in  
684 lagoonal sediments and mussels of Venice, Italy. *Mar. Pollut. Bull.* 26, 501–507.  
685 doi:10.1016/0025-326X(93)90467-X
- 686 Silva, C. a, Madureira, L. a S., 2012. Source correlation of biomarkers in a mangrove ecosystem on  
687 Santa Catarina Island in southern Brazil. *An. Acad. Bras. Cienc.* 84, 589–604.
- 688 Simon, L.N., Raffaelli, D., 2012. Assessing ecosystem effects of small–scale cutting of Cameroon  
689 mangrove forests. *J. Ecol. Nat. Environ.* 4, 126–134. doi:10.5897/JENE11.131
- 690 Skov, M.W., Vannini, M., Shunula, J.P., Hartnoll, R.G., Cannicci, S., 2002. Quantifying the density  
691 of mangrove crabs: Ocypodidae and Grapsidae. *Mar. Biol.* 141, 725–732.
- 692 Sousa, W.P., Dangremond, E.M., 2011. Trophic interactions in coastal and estuarine mangrove  
693 forest ecosystems, in: *Treatise on Estuarine and Coastal Science*,. Academic London,  
694 Waltham, pp. 43–93.
- 695 Spalding, M., Kainuma, M., Collins, L., 2010. *World Atlas of mangroves.* Earthscan 319.
- 696 Sparks, D.L., Page, A.L., Helmke, P.A., Loeppert, R.H., Soltanpour, P.N., Tabatabai, M.A.,  
697 Johnston, C.T., Sumner, M.E., 1996. *Methods of soil analysis. Part 3 - chemical methods.* Soil  
698 Science Society of America Inc., Madison, WI.
- 699 Tam, N.F.Y., Wong, Y.S., 1995. Spatial and temporal variations of heavy metal contamination in  
700 sediments of a mangrove swamp in Hong Kong. *Mar. Pollut. Bull.* 31, 254–261.  
701 doi:10.1016/0025-326X(95)00141-9
- 702 UNEP, 2007. *Mangroves of western and central Africa.* Earthprint, Chicago.
- 703 Vane, C.H., Harrison, I., Kim, A.W., Moss-Hayes, V., Vickers, B.P., Hong, K., 2009. Organic and  
704 metal contamination in surface mangrove sediments of South China. *Mar. Pollut. Bull.* 58,  
705 134–144. doi:10.1016/j.marpolbul.2008.09.024
- 706 Walle, N. Van De, 1989. Rice Politics in Cameroon: State Commitment, Capability, and Urban  
707 Bias. *J. Mod. Afr. Stud.* 27, 579. doi:10.1017/S0022278X00020450
- 708 Wong, S., Tam, N.F.Y., Lan, C.Y., 1997. Mangrove wetlands as wastewater treatment facility : a  
709 field trial. *Hydrobiologia* 352, 49–59.
- 710 Woodroffe, C.D., Rogers, K., McKee, K.L., Lovelock, C.E., Mendelssohn, I. a., Saintilan, N., 2016.  
711 *Mangrove Sedimentation and Response to Relative Sea-Level Rise.* *Ann. Rev. Mar. Sci.* 8,

712 annurev-marine-122414-034025. doi:10.1146/annurev-marine-122414-034025  
713 Yang, J., Zhang, W., Shen, Y., Feng, W., Wang, X., 2007. Monitoring of organochlorine pesticides  
714 using PFU systems in Yunnan lakes and rivers , China. *Chemosphere* 66, 219–225.  
715 doi:10.1016/j.chemosphere.2006.05.055  
716 Yi, Y., Yang, Z., Zhang, S., 2011. Ecological risk assessment of heavy metals in sediment and  
717 human health risk assessment of heavy metals in fishes in the middle and lower reaches of the  
718 Yangtze River basin. *Environ. Pollut.* 159, 2575–2585. doi:10.1016/j.envpol.2011.06.011  
719 Yu, R., Chen, G.Z., Wong, Y.S., Tam, N.F.Y., Lan, C.Y., 1997. Benthic macrofauna of the  
720 mangrove swamp treated with municipal wastewater. *Hydrobiologia* 347, 127–137.  
721 Zaccone, C., Gallipoli, A., Cocozza, C., Trevisan, M., Miano, T.M., 2009. Distribution patterns of  
722 selected PAHs in bulk peat and corresponding humic acids from a Swiss ombrotrophic bog  
723 profile. *Plant Soil* 315, 35–45. doi:10.1007/s11104-008-9775-1  
724 Zhou, F., Guo, H., Hao, Z., 2007. Spatial distribution of heavy metals in Hong Kong’s marine  
725 sediments and their human impacts: A GIS-based chemometric approach. *Mar. Pollut. Bull.*  
726 54, 1372–1384. doi:10.1016/j.marpolbul.2007.05.017

727  
728  
729  
730  
731  
732  
733  
734  
735  
736  
737  
738  
739  
740  
741  
742  
743  
744  
745  
746  
747  
748  
749  
750  
751  
752  
753  
754  
755  
756  
757  
758  
759  
760  
761  
762  
763  
764  
765

729 **Tables**730 Table 1. Total PCB concentrations in sediment samples. Sample size is shown in brackets. Data are  
731 expressed as mean  $\pm$  standard error.

Forest	Belt	Depth (cm)	PCB tot ( $\mu\text{g kg}^{-1}$ )
Bois des singes	<i>Rhizophora</i>	10 (n=7)	6.2 $\pm$ 1.9
		20 (n=7)	6.3 $\pm$ 1.8
Wouri Bridge	<i>Avicennia</i>	10 (n=3)	6.3 $\pm$ 0.4
		20 (n=3)	5.0 $\pm$ 3
	<i>Pandanus</i>	10 (n=3)	2.6 $\pm$ 1.2
		20 (n=3)	-
	<i>Rhizophora</i>	10 (n=3)	5.3 $\pm$ 1.5
		20 (n=3)	4.0 $\pm$ 1.2

1732  
1733  
1734 Table 2. p-p'DDE and DEHP concentrations in sediment samples. Sample size is shown in  
1735 brackets. Data are expressed as mean  $\pm$  standard error.

Forest	Belt	Depth (cm)	p-p DDE ( $\mu\text{g kg}^{-1}$ )	DEHP ( $\mu\text{g kg}^{-1}$ )
Bois des singes	<i>Rhizophora</i>	10 (n=7)	30 $\pm$ 0.2	750 $\pm$ 110
		20 (n=7)	10 $\pm$ 0.1	540 $\pm$ 90
Wouri Bridge	<i>Avicennia</i>	10 (n=3)	-	710 $\pm$ 370
		20 (n=3)	-	530 $\pm$ 110
	<i>Pandanus</i>	10 (n=3)	-	380 $\pm$ 40
		20 (n=3)	-	1008 $\pm$ 390
	<i>Rhizophora</i>	10 (n=3)	40 $\pm$ 0.07	750 $\pm$ 290
		20 (n=3)	40 $\pm$ 0.07	970 $\pm$ 60

1736  
1737 Table 3. Stanol contamination index: percentage of coprostan-3-ol and 5  $\beta$ -cholestan-3 $\alpha$ -ol on total  
1738 sterols, calculated for the different depths in each belt of the two forests. High levels of  
1739 contaminants (according with Froehner et al., 2009) are shown in bold.

Samples	Site	Depth	Belt	% (coprostan-3-ol + 5 $\beta$ -cholestan-3 $\alpha$ -ol )/total stanols
A1PW10	Wouri Bridge	10	<i>Avicennia</i>	<b>43.9</b>
A1PW20	Wouri Bridge	20	<i>Avicennia</i>	0.1
P1PW10	Wouri Bridge	10	<i>Pandanus</i>	9.8
P1PW20	Wouri Bridge	20	<i>Pandanus</i>	0.3
P2PW10	Wouri Bridge	10	<i>Pandanus</i>	<b>47.8</b>
P2PW20	Wouri Bridge	10	<i>Pandanus</i>	5.4
R1PW10	Wouri Bridge	10	<i>Rhizophora</i>	6.3
R1PW20	Wouri Bridge	20	<i>Rhizophora</i>	7.1
R2PW10	Wouri Bridge	10	<i>Rhizophora</i>	<b>44.5</b>
R2PW20	Wouri Bridge	20	<i>Rhizophora</i>	3.3
R3PW10	Wouri Bridge	10	<i>Rhizophora</i>	<b>67.4</b>
R3PW20	Wouri Bridge	20	<i>Rhizophora</i>	9.0
R1BS10	Bois des singes	10	<i>Rhizophora</i>	24.0
R1BS20	Bois des singes	20	<i>Rhizophora</i>	3.7
R2BS10	Bois des singes	10	<i>Rhizophora</i>	11.9
R2BS20	Bois des singes	20	<i>Rhizophora</i>	21.6
R3BS10	Bois des singes	10	<i>Rhizophora</i>	<b>43.1</b>
R3BS20	Bois des singes	20	<i>Rhizophora</i>	<b>39.1</b>

R4BS10	Bois des singes	10	<i>Rhizophora</i>	23.0
R4BS20	Bois des singes	20	<i>Rhizophora</i>	<b>25.6</b>
R5BS10	Bois des singes	10	<i>Rhizophora</i>	21.5
R5BS20	Bois des singes	20	<i>Rhizophora</i>	<b>35.6</b>
R6BS10	Bois des singes	10	<i>Rhizophora</i>	<b>26.0</b>
R6BS20	Bois des singes	20	<i>Rhizophora</i>	14.1
R7BS10	Bois des singes	10	<i>Rhizophora</i>	<b>68.2</b>
R7BS20	Bois des singes	20	<i>Rhizophora</i>	18.8

Ratio values that represent detection of contamination are shown in bold. BS = Bois des Singes; PW=Wouri Bridge.

Table 4. Sediment water temperature, pH and Conductivity and pore water Total Nitrogen (TN) and Organic Carbon (TOC) recorded in each plot during macrobenthos surveys. Data are expressed as mean  $\pm$  standard error.

Forest	Belt	Temperature (°C)	pH	Conductivity (mV)	Tot N (%)	Tot OC (%)
Bois des singes	<i>Rhizophora</i>	27.35 $\pm$ 0.3	6.29 $\pm$ 0.06	16.13 $\pm$ 2.9	0.24 $\pm$ 0.04	4.09 $\pm$ 0.49
Wouri Bridge	<i>Avicennia</i>	26.28 $\pm$ 0.4	6.25 $\pm$ 0.04	20.83 $\pm$ 2.3	0.28 $\pm$ 0.09	4.55 $\pm$ 1.38
	<i>Pandanus</i>	26.9 $\pm$ 0.4	6.30 $\pm$ 0.03	18.08 $\pm$ 1.6	0.33 $\pm$ 0.2	5.65 $\pm$ 0.19
	<i>Rhizophora</i>	26.18 $\pm$ 0.06	6.29 $\pm$ 0.07	19 $\pm$ 2	0.38 $\pm$ 0.1	6.01 $\pm$ 1.25

Table 5. Test for relationships between sterols, metals, PAHs and C/N with macrobenthos distribution, using permutational multiple regression analysis (DISTLM). AICc: coefficient of regression, SS: sum of squares, F: value of pseudo and its significance p (% Var: percentage of variance explained by each single variable, and % Cumul: cumulative percentage of variance explained, Res. df: residual degrees of freedom). In bold the variable statistically significant related with macrobenthos distribution.

Variable	AICc	SS(trace)	Pseudo-F	p	% Var.	%Cumul.	Res.df
+ coprostan-3-ol	0.012	113.38	0.29	0.7352	0.001	0.012	24
+ <b>5 <math>\beta</math>-cholestan-3<math>\alpha</math>-ol</b>	<b>0.219</b>	<b>1978.7</b>	<b>6.12</b>	<b>0.0105</b>	<b>0.208</b>	<b>0.219</b>	<b>23</b>
+cholesterol	0.291	678.86	2.21	0.1209	0.071	0.291	22
+5 $\alpha$ -cholestan-3 $\beta$ -ol	0.314	219.05	0.70	0.4644	0.023	0.314	21
+As	<b>0.434</b>	<b>1151.3</b>	<b>4.27</b>	<b>0.0316</b>	<b>0.121</b>	<b>0.434</b>	<b>20</b>
+Se	<b>0.587</b>	<b>1458.1</b>	<b>7.05</b>	<b>0.0073</b>	<b>0.153</b>	<b>0.587</b>	<b>19</b>
+Mo	0.604	155.92	0.74	0.471	0.016	0.604	18
+Cd	0.625	197.66	0.94	0.3786	0.021	0.625	17

	+Sn	0.637	119.91	0.55	0.5553	0.013	0.637	16
1	+Sb	0.669	300.56	1.43	0.241	0.032	0.669	15
2								
3	<b>+Cr</b>	<b>0.751</b>	<b>785.68</b>	<b>4.64</b>	<b>0.0322</b>	<b>0.082</b>	<b>0.751</b>	<b>14</b>
4	+Cu	0.804	508.87	3.55	0.0613	0.053	0.804	13
5	+Mn	0.809	45.498	0.30	0.7313	0.005	0.809	12
6								
7	<b>+Zn</b>	<b>0.885</b>	<b>717.9</b>	<b>7.18</b>	<b>0.0095</b>	<b>0.075</b>	<b>0.885</b>	<b>11</b>
8	+phenanthrene	0.900	149.18	1.57	0.2309	0.016	0.900	10
9	+fluoranthene	0.901	6.8011	0.06	0.8834	0.001	0.901	9
10	+ benzo[a] anthracene	0.918	166.41	1.71	0.2141	0.017	0.918	8
11	+ benzo							
12	[jbc]fluoranthene	0.922	38.665	0.37	0.71	0.004	0.922	7
13	+ benzo [a] pyrene	0.922	-3.598	-0.03	0.9607	0.000	0.922	6
14	+DEHP	0.968	85.979	1.13	0.3724	0.009	0.968	4
15	+p-p'DDE	0.984	149.9	2.91	0.1425	0.016	0.984	3
16								
17	+C/N	0.990	56.641	1.16	0.365	0.006	0.990	2

19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

766

**Figures and Figure Captions**

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

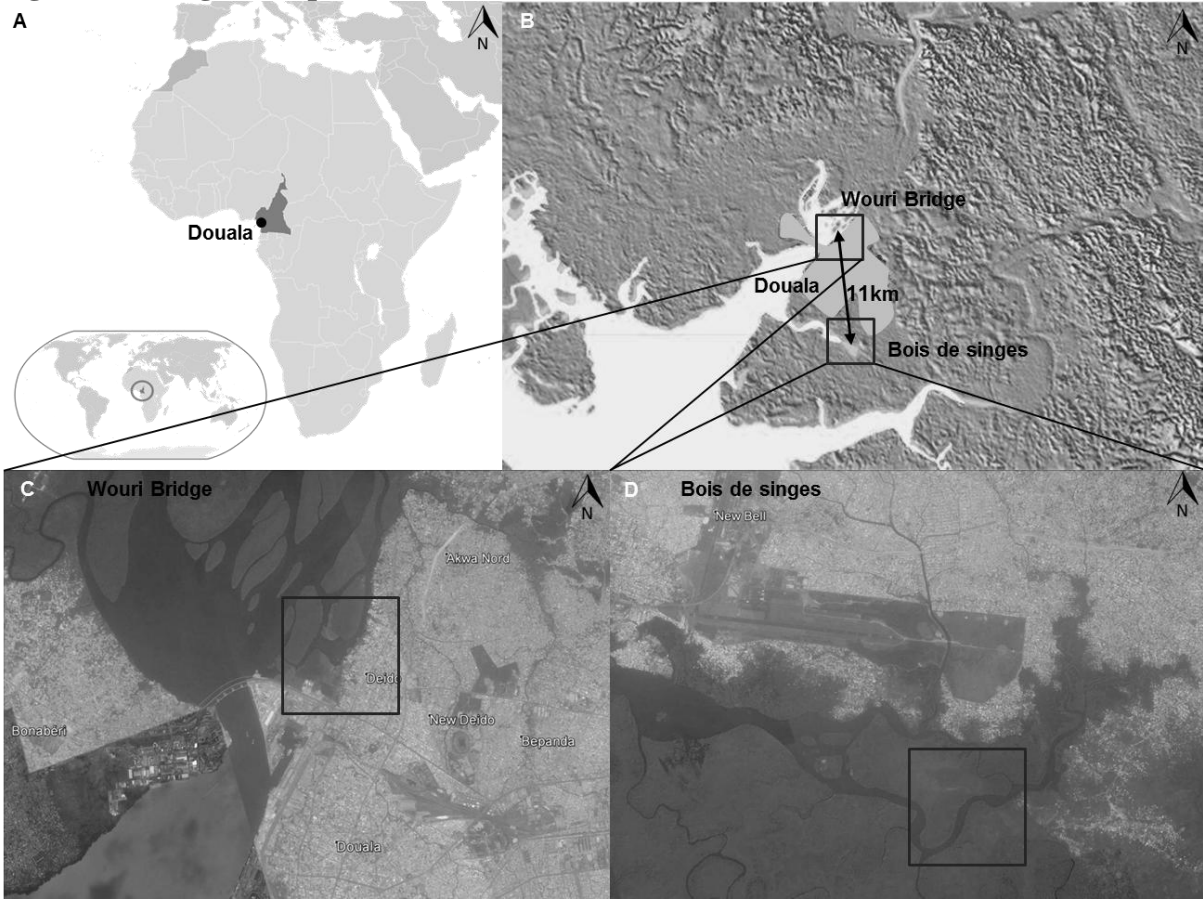
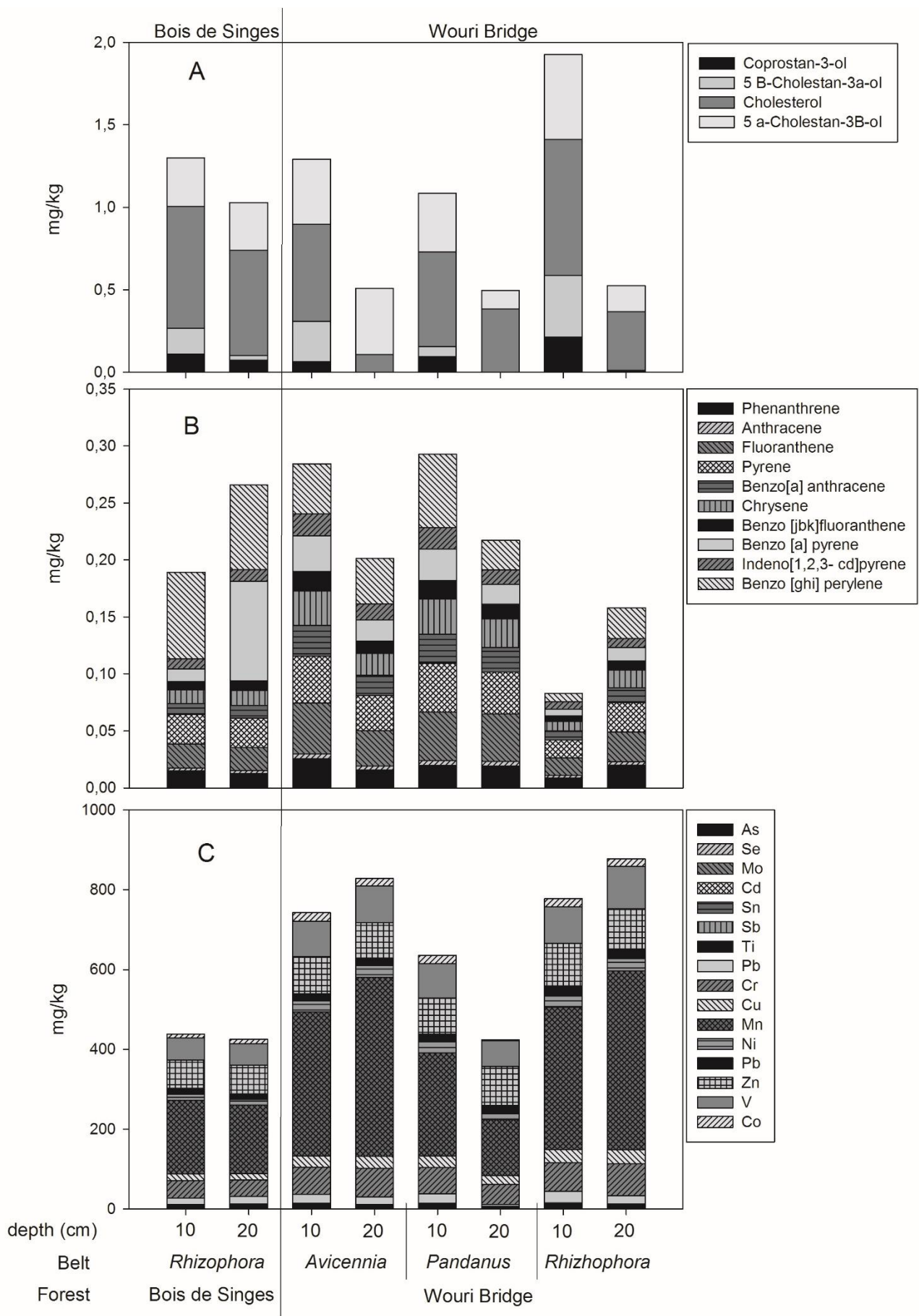


Figure 1. Study sites. A) Continental overview, B) Woury Estuary where Douala is located and C) Wouri Bridge mangrove forest (WB) and D) Bois de singes mangrove forest (BS). Black squares indicate the exact location where the study was carried on (Image source: Google Earth).

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65





773 Figure 2. Average concentrations of Sterols (A), PAHs (B) and metals (C), detected in sediment  
774 samples. Data are shown according to vegetation belt (*Avicennia* sp., *Pandanus* sp., *Rhizophora* sp.)  
775 and sampling depth on the x-axis. Surface sediment upper layer (0-10 cm) and core samples from  
776 11 to 20 cm.  
777

6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

778

779

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

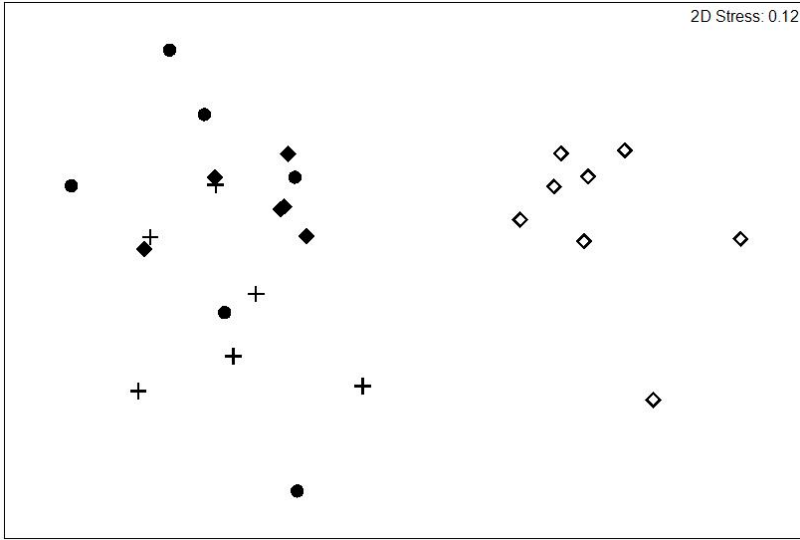


Figure 3. Non-metric multidimensional scaling ordination showing the patterns of distribution of macrobenthic species in the two study forests.  $\diamond$  Bois des Singes *Rhizophora*;  $\bullet$  Wouri Bridge *Avicennia*; + Wouri Bridge *Pandanus*;  $\blacklozenge$  Wouri Bridge *Rhizophora*.

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

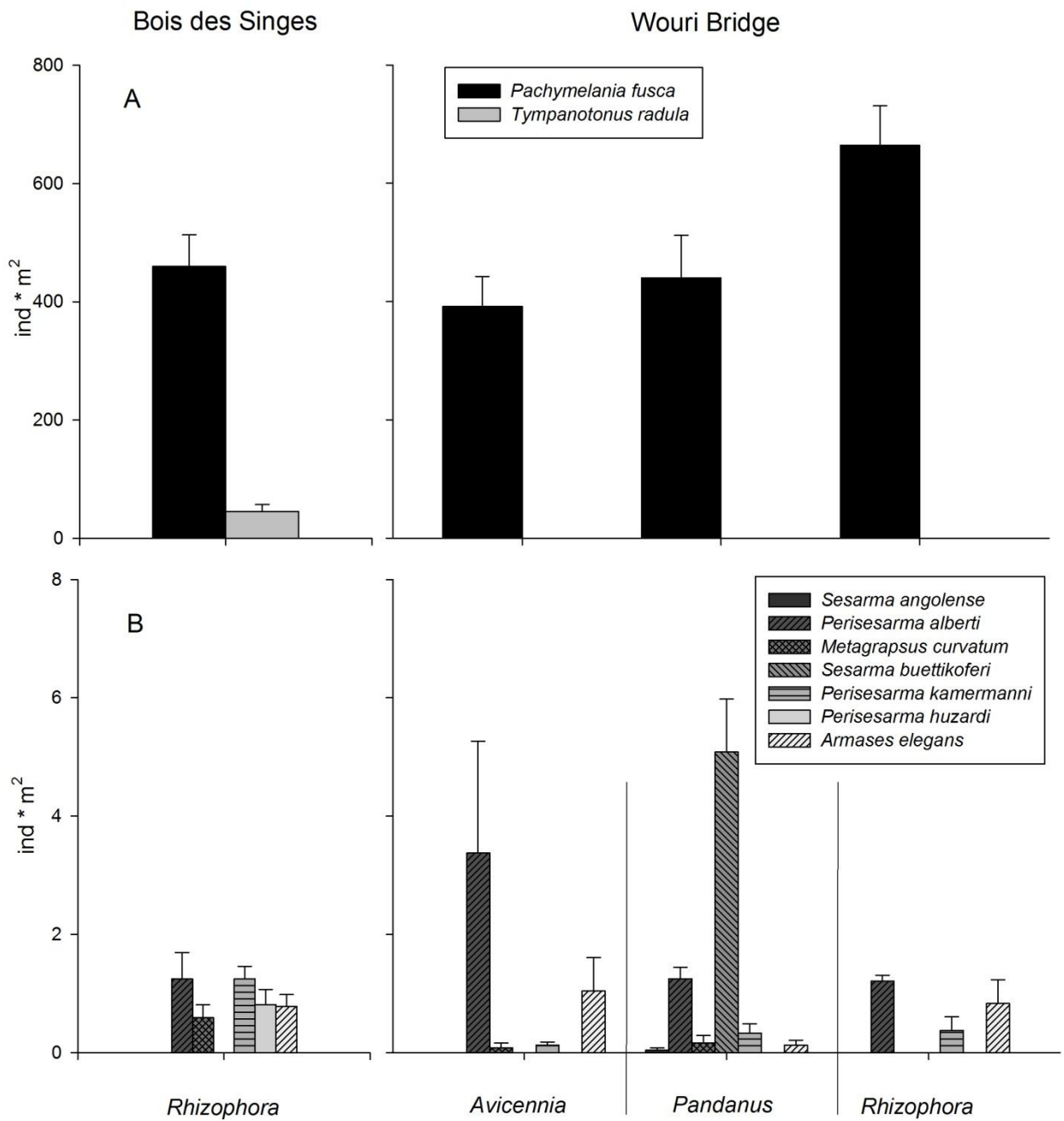


Figure 4. Densities of mollusc (A) and crab (B) species in the study sites. Values are expressed as mean ± SE.

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

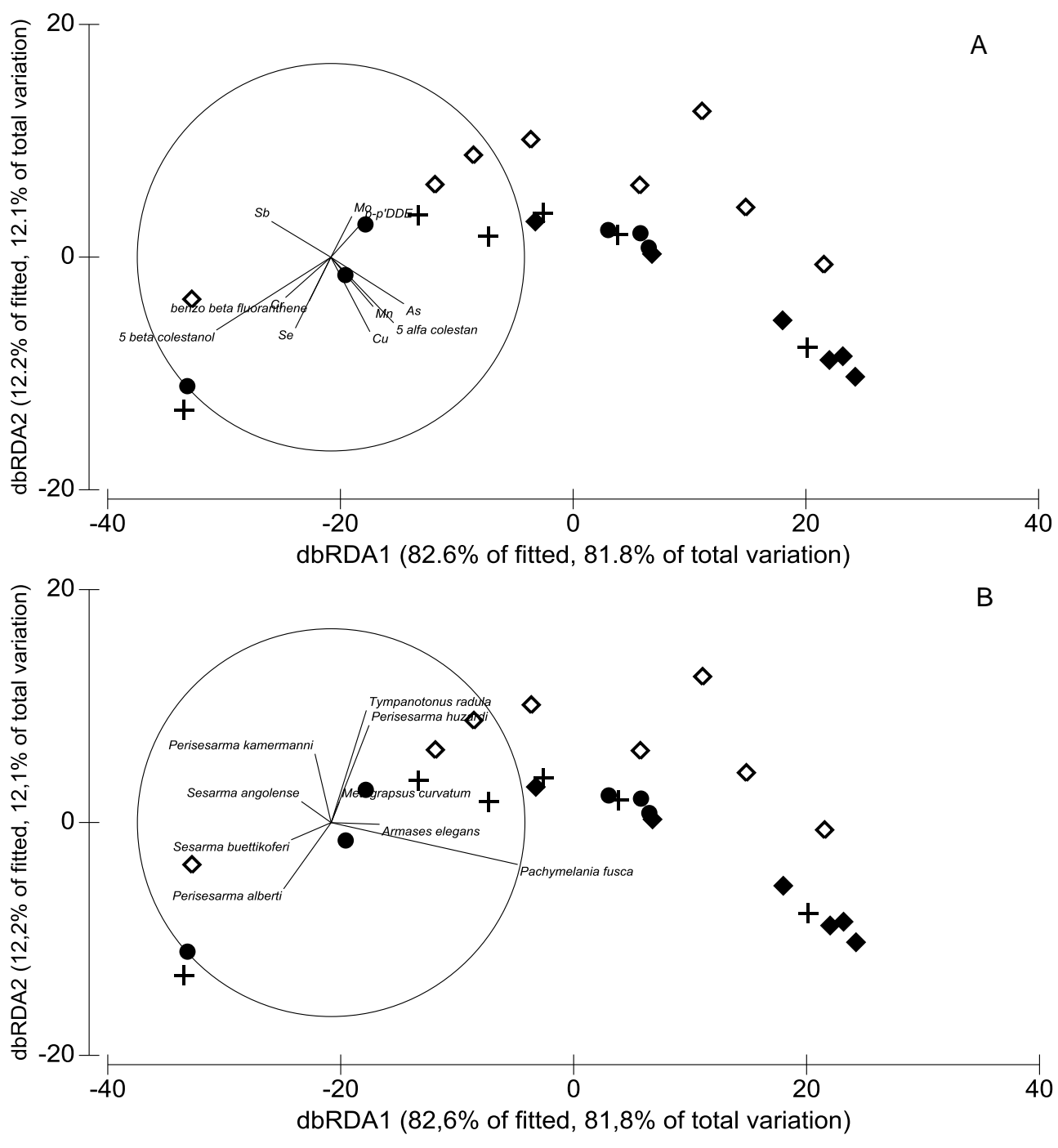


Figure 5. Distance-based redundancy analysis plots (dbRDA) of macrobenthos distribution across Wouri Bridge (*Avicennia* belt (●), *Pandanus* belt (+) and *Rhizophora* belt (◆)) and Bois des Singes (◇), in accordance with the contaminants found in the sediment core of each belt. Vectors correspond to environmental variables (A) and species (B). Length and direction of the vectors indicate the strength of the correlation between the variable and ordination axis given the other variables in the model. The radius of the circle denotes a correlation of 1.

**Supplementary Data**

[Click here to download Supplementary Data: Fusi et al SM.docx](#)